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Tissue Chips in Space

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Abstract

The International Space Station (ISS) U.S. National Laboratory in collaboration with the National Center for Advancing Translational Sciences (NCATS) and the National Institute of Biomedical Imaging and Bioengineering (NIBIB) at the National Institutes of Health (NIH) developed the "Tissue Chips in Space" initiative to promote and fund research into human physiology and disease in low Earth orbit (LEO) that will translate into advancements in Earth-based medicine. The Tissue Chips in Space initiative is part of NIH's larger Tissue Chip Program [1] that aims to develop bioengineered devices to improve the complex and expensive process of predicting whether drugs will be safe and effective or toxic in humans. These bioengineered devices, referred to as microphysiological systems, "tissue chips," or "organs-on-a-chip," leverage recent advances in cell biology, tissue engineering, and microfabrication to accurately model human organ tissues in *in vitro* platforms. These systems offer promising solutions for modelling human physiology and disease pathology for applications in areas where traditional cell culture and animal models fall short. This report provides an overview of the Tissue Chips in Space initiative, an update on the its current status, and a discussion of its potential long-term benefits.

Keywords: tissue chips, micro physiological systems, international space station, disease modeling, space flight effect

Acronyms/Abbreviations

Blood brain barrier (BBB) Central nervous system (CNS) Children's Hospital of Philadelphia (CHOP) Defense Advanced Research Projects Agency (DARPA) Induced pluripotent stem cells (iPSC) International Space Station (ISS) Massachusetts Institute of Technology (MIT) National Center for Advancing Translational Sciences (NCATS) National Institute of Biomedical Imaging and Bioengineering (NIBIB) National Institutes of Health (NIH) National Aeronautics and Space Administration (NASA)

Post-traumatic osteoarthritis (PTOA)

SpaceX (SpX)

University of California, San Francisco (UCSF)

University of Florida (UF) University of Washington (UW) U.S. Food and Drug Administration (FDA)

1. Introduction

1.1 The Collaborators

To expand the research opportunities available onboard the International Space Station (ISS), in 2005, Congress designated the U.S. portion of the ISS as a U.S. National Laboratory, facilitating access to spacebased research and development for a broad range of commercial, academic, and government users. The ISS U.S. National Laboratory is responsible for managing all non-NASA research, and all ISS National Laboratory-sponsored investigations must have the capacity to utilize the ISS for Earth benefits. By engaging with other government agencies in the public sector and with universities and companies in the private sector, the ISS National Laboratory promotes and brokers a diverse range of research in life sciences, physical sciences, remote sensing, technology development, and education [2].

The National Center for Advancing Translational Sciences (NCATS) at the National Institutes of Health (NIH) was officially established in fiscal year 2012 to transform the translational science process so that new treatments and cures for disease could be delivered to patients faster. NCATS, one of 27 Institutes and Centers at NIH, strives to develop innovations to reduce, remove, or bypass costly and time-consuming bottlenecks in the translational research pipeline in an effort to speed the delivery of new drugs, diagnostics, and medical devices to patients [3].

The National Institute of Biomedical Imaging and Bioengineering (NIBIB) is authorized by the National Institute of Biomedical Imaging and Bioengineering Establishment Act H.R. 1795, which was signed into law on December 29, 2000. NIBIB's mission is to improve health by leading the development and accelerating the application of biomedical technologies. The institute is committed to integrating the physical and engineering sciences with the life sciences to advance basic research and medical care. [4].

1.2 The Problem

Human physiology and disease pathology involve complex interactions between different cells. extracellular matrices, cell signalling molecules, and other factors. These interactions occur both in specific microenvironments as well as on the larger systemic level. However, both healthy and disease modalities have traditionally been studied using reductionist approaches that include the use of two-dimensional cell cultures that are often void of their native extracellular environment and lack the three-dimensionality that cells maintain in vivo. In part, these reductionist approaches have been used to take advantage of advances in robotics and the adoption of high-throughput screening programs. While these simplified approaches are useful in some cases, such as for validating target proteins or pathways and identifying hit compounds for development, they risk providing results that are outside the context of the larger system that human diseases actually exist in.

On the whole systems side, non-human surrogates are often used to study disease pathology and validate therapeutic interventions. These surrogates include rodents, dogs, nonhuman primates, zebra fish, and larger mammals such as pigs. Although these animals provide many advantages for research, including the ability to model systemic responses in complex organisms, be subjects for genetic editing tools, and reduce the risk for first-in-human tests, animals have different physiological and genetic responses, and there is a push for development of novel human systems that faithfully model human physiology and reduce animal use in research [5-7].

Recent studies have shown that, on average, across all indications, there is less than a 14% chance that a drug entering clinical trials will succeed in receiving U.S. Food and Drug Administration (FDA) approval [8]. The primary source of drug failure is due to an inability to demonstrate efficacy [9]. Another main reason for failure is safety, which is also monitored post-approval and can result in the removal of a therapeutic from the market. While a number of factors can contribute to the failure of a clinical candidate, it is well-recognized that the lack of relevant and appropriate preclinical models is a factor in many cases.

2. Tissue Chips

With the recent advancements in cell biology, tissue engineering, and microfabrication, there has been a multidisciplinary approach to building systems that are more human-relevant for use in studying disease pathology and validating therapeutic efficacy and safety. As part of this effort, many groups have developed tissue chip platforms that utilize 3D human multicellular complexes. These complexes are engineered into a structural environment that attempts to replicate the native in vivo environment. Such tools provide a more useful model than traditional 2D cultures, and through the use of human cells (healthy, diseased, or engineered) can replicate patient-specific pathology and open up avenues for precision and personalized medicine approaches [10].

To date, numerous studies have utilized tissue chip platforms. Tissue chips have been used to recapitulate functional tissues from organs such as bone, brain, gut, liver, lung, skin, and others [11-15] and to model diseases [10], including rare diseases [16] such as Progeria [17] and Barth Syndrome [18]. Tissue chips have also been utilized to preform pharmacokinetic testing as well as efficacy and toxicity screening [19-21]. In addition, tissue chip platforms have been used to study cancer biology [22,23], and much progress has been made to link multiple tissue chips together to study interactions between different organs and model systemic responses [24-26].

The development of tissue chip technology has been supported by the NIH Tissue Chip Program, which issued 19 awards in 2012 to investigators to develop human tissue chips that would provide better predictions of drug safety and efficacy than current models. The program ran alongside a sister program supported by the Defense Advanced Research Projects Agency (DARPA) which focused on funding the development of 10 linked organ systems that would model system effects of disease and be useful for drug evaluation. The NIH Tissue Chip Program has now expanded to over 40 teams supported by more than 15 NIH Institutes or Centers. The program is heavily collaborative with other government agencies such as the FDA and commercial partners from the pharmaceutical industry in order to expedite the translation of discoveries into therapeutics. In 2016, the ISS National Laboratory and NCATS entered into a collaboration to create the Tissue Chips in Space initiative, and in 2018, NIBIB joined the initiative [27].

3. Tissue Chips in Space

It is well documented that microgravity accelerates changes in human physiology, such as muscle wasting, osteoporosis, reduced cardiopulmonary function, and altered immune response, among others, and in many instances, these changes directly correlate to disease pathology on Earth [28-31]. By utilizing tissue chips containing human cells on the ISS National Laboratory, disease pathologies that might take years to produce on Earth are accelerated and can be studied on an expedited time frame.

To date, the Tissue Chips in Space initiative has funded nine projects, five of which were awarded from NIH RFA-TR-16-019 and four of which were awarded from NIH RFA-TR-18-001 (see Table 1). Additionally, as many of these changes are reversible upon return to Earth, these studies are suitable for developing drug targets. The awards, which are issued as cooperative agreements managed by NIH, require the teams to adhere to strict timelines and meet quantitative milestones in order to progress. The initiative provides the opportunity for each awarded team to launch two separate experiments to the ISS, contingent on successful completion of each project's milestones. The first phase of each project is designed to validate the disease model of study, and the second is to test novel therapeutics in the model. Each awarded team works with an Implementation Partner, a commercial company which is responsible for payload development and flight logistics such as manifesting and safety certifications.

As noted previously, the goal of the Tissue Chips in Space initiative is to use tissue chip platforms and the unique microgravity environment of the ISS to develop models of human disease, with the ultimate goal of expediting the discovery of therapeutics for people on Earth. In order to accomplish this goal, awarded teams leverage previous knowledge regarding the effects of spaceflight on the human body and how those effects translate to human diseases on Earth (see Table 1 for a summary of projects funded through the Tissue Chips in Space Program). For example, a team from the University of California, San Francisco (UCSF) is using the known dysregulation of the immune system and inflammatory responses associated with spaceflight as a surrogate model for Earth-based immunosenescence. The project is specifically investigating microgravityinduced aging of the immune system (generated through

simulated microgravity and spaceflight) and its role in tissue-specific healing and regeneration.

A team of researchers from the Massachusetts Institute of Technology (MIT) is exploring putative therapeutics to treat post-traumatic osteoarthritis (PTOA). Utilizing a tissue chip system comprised of human cartilage, bone, and synovium that is challenged with inflammatory cytokines and an acute impact injury, the team will validate the system as an appropriate model for PTOA (see Fig. 1). Once validated, the team will use the model to test therapeutic options and monitor their effectiveness through the use of intracellular and extracellular biomarkers.



Fig. 1. Canadian Space Agency astronaut David Saint-Jacques works with the Cartilage-Bone-Synovium Tissue Chips in Space investigation. Image Credit: NASA

A University of Washington (UW) team is leveraging the ISS environment to study proximal tubule proteinuria and distal tubule kidney stone formation (see Fig. 2). Kidney dysfunction can result in serious health problems such as proteinuria, osteoporosis, and kidney stone formation in patients on Earth. However, disease progression is often slow and difficult to model *in vivo* in terrestrial studies. Using a human cell-derived kidney tissue chip system onboard the ISS, the team hopes to accelerate the onset of the disease states to produce a system that will allow for rapid translation to Earth-based therapeutics.



Fig. 2. NASA astronaut Anne McClain works inside the Life Sciences Glovebox with the Tissue Chips in Space Kidney Cells investigation. Image Credit: NASA

The blood brain barrier (BBB) is a critically vital component of the body that serves as a gatekeeper between the circulating blood in the brain and extracellular fluid in the central nervous system (CNS). A model that allows monitoring of the dysregulation of the BBB could provide applications for studying neurological disorders as well as the transport of drugs and toxins to the CNS. To this effect, a team of researchers from Emulate is developing an automated BBB tissue chip platform derived from human cells for use both on the ground and on the ISS (see Fig. 3).



Fig. 3. The Organs-On-Chips as a Platform for Studying Effects of Microgravity on Human Physiology

investigation is shown near the Powered Ascent Utility Locker. Image Credit: NASA

The innate immune response of humans allows for the recruitment of immune cells to an infected organ. A research team from the Children's Hospital of Philadelphia (CHOP) is developing a human airway tissue chip and connecting it to a bone marrow tissue chip. Using this interconnected system, the team can infect the airway chip and monitor the recruitment of neutrophils from the bone marrow as a model of innate response. Capitalizing on known spaceflight-induced changes associated with dysregulation of the immune system, this ISS project will serve as a model for a compromised immune system (see Fig. 4).



Fig. 4: The Lung Host Defence in Microgravity Chips in Space investigation hardware floats in the Destiny module of the International Space Station. Image Credit: NASA

Sarcopenia is characterized by the loss of skeletal muscle mass and function and commonly affects older adults. It is well documented that humans exposed to long-term spaceflight experience muscle wasting. A team from the University of Florida (UF) aims to use the accelerated muscle wasting environment of the ISS in combination with a human muscle tissue chip as a model for sarcopenia in terrestrial settings. Through the use of cells from different types of patients (human myocytes isolated from young, healthy and older, sedentary volunteers), the team will monitor the progression of the sarcopenia phenotype on a time scale not possible in terrestrial settings.

A team from Stanford University is utilizing human induced pluripotent stem cells (iPSC) from healthy patients to fabricate engineered heart tissue platforms for use on the ISS and on the ground. By taking advantage of microgravity-induced weakening of the heart muscle, the team will validate the ISS platform as a tool to model ischemic cardiomyopathy in humans on Earth. Once the model is validated, the team will use the platform to screen for potential drug candidates to treat patients. Similarly, in another project from the University of Washington (UW), the team will use an engineered heart tissue chip to study aspects of cardiomyopathy associated with human health on Earth and in space. The project will utilize a novel magnetometer-based motion sensor to allow for automated real-time continuous functional readouts of the engineered heart tissues. The team will ultimately test pharmaceuticals and mechanical stimulations as potential therapeutic interventions.

Lastly, in a second project from Emulate, the team will use plug-and-play technologies to adapt their automated platform developed for their BBB ISS experiment to study dysregulation in the gut. The team will challenge the system with an infection and study the innate response and probiotic-induced response to the infection.

4. Conclusion

The work conducted through the Tissue Chips in Space initiative aims to leverage tissue chip technology onboard the ISS National Laboratory to lead to expedited discovery of therapeutics to treat patients on Earth. In addition, several ancillary benefits have already occurred as a result of the initiative, including the development of miniaturized automated systems for platform support. Sending tissue chips to the ISS requires the development of hardware that is automated and reliable with a small footprint. For example, in preparation for their spaceflight experiment, the University of Washington team, in collaboration with Implementation Partner Bioserve Space Technologies (see Table 1 for other implementation partners involved in the Tissue Chips in Space initiative), was able to reduce their tissue chip system from a 1,350-L volume system to a 45-L volume system [50]. In addition, tissue chip hardware for spaceflight must be designed to survive launch, operation onboard the ISS, and splashdown and must be capable of producing robust scientific results. The advancements that result from translating terrestrial systems to spaceflight systems are necessary steps in making tissue chip technology adaptable for wide-spread use in ground-based applications and to increase accessibility for greater numbers of researchers on Earth. One of the current limiting factors in the wide-spread adoption of tissue chip systems is the requirement for extensive hands-on interactions with the experiment. Through this program, this limitation is being addressed by development of automated and simplified systems such as these ones designed for spaceflight that allow for plug-and-play use. Overall, the need for these experiments to run in multiple terrestrial locations, as well as in orbit, without experimental confounders has pushed the biological and technical validation of the platforms, which translates into benefit for the whole field. As a result of the Tissue Chips in Space initiative, several commercial companies

and other government agencies have begun projects that may utilize tissue chips on the ISS in order to address their own organizational mandates, further expanding and validating the use of tissue chip technology. Finally, while the primary purpose of the Tissue Chips in Space initiative is to translate results to benefit life on Earth, it is straightforward to imagine application of this technology to understand and mitigate the risks to human health posed by long-duration spaceflight.

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Tables

Table 1. Summary of projects funded through the Tissue Chips in Space initiative. Principle Investigator (PI) refers to the PI listed as the Contact PI on the NIH grant. Awardee Organization is the institution awarded the management of the grant. UCSF = University of California at San Francisco, MIT = Massachusetts Institute of Technology, UW = University of Washington, CHOP = Children's Hospital of Philadelphia, SpX = SpaceX. Grant # is the current NIH Project Number associate with the grant (search https://projectreporter.nih.gov/reporter.cfm with the project number for additional details about each project). Spaceflight Effect is the known spaceflight effects the teams are looking to utilize in their models. Translation is the potential Earth-based therapeutic applications the projects are targeting. First Flight and Second Flight are the targeted launch vehicles for the given payloads. One project was launched on SpX-16 in 2018, and four were launched on SpX-17 in 2019. The remaining projects' launch vehicles and dates are currently to be determined (TBD).

Principle Investigator (Awardee Organization)	Implementation Partner	Grant #	Spaceflight Effect	Translation	First Flight (year)	Second Flight (year)
Sonja Schrepfer (UCSF)	Bioserve Space Technologies	UG3TR002192	Dysregulated immune system [32-34], inflammatory response [35,36]	Immunological Senescences [37,38], Adaptive Immunity [39], Impaired Myocardial Regeneration [40]	SpX-16 (2018)	TBD (2020)
Alan Grodzinsky (MIT)	Techshot	UG3TR002186	Musculoskeletal injuries and disease [41-44]	Post-traumatic osteoarthritis [45]	SpX-17 (2019)	TBD (2021)
Jonathan Himmelfarb (UW)	Bioserve Space Technologies	UG3TR002178	Dysregulation of kidney function [46-48]	Proteinuric chronic kidney disease [49,50]	SpX-17 (2019)	TBD (2021)
Christopher Hinojosa (Emulate)	Space Tango	UG3TR002188	Immune system deterioration [51], blood brain barrier integrity [52]	Multiple sclerosis [53], epilepsy [54]	SpX-17 (2019)	TBD (2021)
Scott Worthen (CHOP)	Space Tango	UG3TR002198	Dysregulated immune system [32-34], poor myeloid cell mobilization [55-56]	Treatment of airway infections [57]	SpX-17 (2019)	TBD (2021)
Siobhan Malany (UF)	Space Tango	UG3TR002598	Muscle atrophy [58-60]	Sarcopenia [61,62]	TBD (2020)	TBD (2022)
Joseph Wu (Stanford University)	Bioserve Space Technologies	UG3TR002588	Cardiomyopathy [63-65]	Cardiovascular disease [66,67]	TBD (2020)	TBD (2022)
Deok-Ho Kim (UW)	Bioserve Space Technologies	UG3TR028094	Cardiomyopathy [63-65,68]	Cardiac atrophy [69], cardiac arrhythmias [70]	TBD (2020)	TBD (2022)
Christopher Hinojosa (Emulate)	Space Tango	UG3TR002595	Gastrointestinal dysfunction [71,72]	Gut microbiome [73,74]	TBD (2020)	TBD (2022)

References

[1] https://ncats.nih.gov/tissuechip (accessed 30.08.19).

[2] https://www.issnationallab.org (accessed 30.08.19).

[3] https://ncats.nih.gov/about/center (accessed 30.08.19).

[4] https://www.nibib.nih.gov/about-nibib/mission (accessed 30.08.19).

[5] Hartung T. Thoughts on limitations of animal models. Parkinsonism Relat Disord. 2008;14 Suppl 2:S81-3.

[6] Gauthier C, Tavernier G, Trochu JN, Leblais V, Laurent K, Langin D, Escande D, Le Marec H. Interspecies differences in the cardiac negative inotropic effects of beta(3)-adrenoceptor agonists. J Pharmacol Exp Ther. 1999 Aug;290(2):687-93.

[7] Teixidó E, Krupp E, Amberg A, Czich A, Scholz S. Species-specificdevelopmental toxicity in rats and rabbits: Generation of a reference compoundlist for development of alternative testing approaches. Reprod Toxicol. 2018 Mar;76:93-102.

[8] Wong CH, Siah KW, Lo AW. Estimation of clinical trial success rates and related parameters. Biostatistics. 2019 Apr 1;20(2):273-286.

[9] Fogel DB. Factors associated with clinical trials that fail and opportunities for improving the likelihood of success: A review. Contemp Clin Trials Commun. 2018;11:156–164. Published 2018 Aug 7.

[10] Low LA, Tagle DA. Tissue chips - innovative tools for drug development and disease modeling. Lab Chip. 2017 Sep 12;17(18):3026-3036.

[11] Phan DT, Bender RHF, Andrejecsk JW, et al. Blood-brain barrier-on-a-chip: Microphysiological systems that capture the complexity of the blood-central nervous system interface. Exp Biol Med (Maywood). 2017;242(17):1669–1678.

[12] Chin E, Goh E. Blood-brain barrier on a chip. Methods Cell Biol. 2018;146:159-182.

[13] Tsamandouras N, Chen WLK, Edington CD, Stokes CL, Griffith LG, Cirit M. Integrated Gut and Liver Microphysiological Systems for Quantitative In Vitro Pharmacokinetic Studies. AAPS J. 2017 Sep;19(5):1499-1512.

[14] Beckwitt CH, Clark AM, Wheeler S, Taylor DL, Stolz DB, Griffith L, Wells A. Liver 'organ on a chip'. Exp Cell Res. 2018 Feb 1;363(1):15-25.

[15] Huh DD. A human breathing lung-on-a-chip. Ann Am Thorac Soc. 2015 Mar;12 Suppl 1:S42-4

[16] Low LA, Tagle DA. Microphysiological Systems (Tissue Chips) and their Utility for Rare Disease Research. Adv Exp Med Biol. 2017;1031:405-415.

[17] Ribas J, Zhang YS, Pitrez PR, Leijten J, Miscuglio M, Rouwkema J, Dokmeci MR, Nissan X, Ferreira L, Khademhosseini A. Biomechanical Strain Exacerbates Inflammation on a Progeria-on-a-Chip Model. Small. 2017 Apr;13(15).

[18] Wang G, McCain ML, Yang L, He A, Pasqualini FS, Agarwal A, Yuan H, Jiang D, Zhang D, Zangi L, Geva J, Roberts AE, Ma Q, Ding J, Chen J, Wang DZ, Li K, Wang J, Wanders RJ, Kulik W, Vaz FM, Laflamme MA, Murry CE, Chien KR, Kelley RI, Church GM, Parker KK, Pu WT. Modeling the mitochondrial cardiomyopathy of Barth syndrome with induced pluripotent stem cell and heart-on-chip technologies. Nat Med. 2014 Jun;20(6):616-23.

[19] Skardal A, Shupe T, Atala A. Organoid-on-achip and body-on-a-chip systems for drug screening and disease modeling. Drug Discov Today. 2016 Sep;21(9):1399-1411.

[20] Cirit M, Stokes CL. Maximizing the impact of microphysiological systems with in vitro-in vivo translation. Lab Chip. 2018 Jun 26;18(13):1831-1837.

[21] Truskey GA. Human Microphysiological Systems and Organoids as in Vitro Models for Toxicological Studies. Front Public Health. 2018 Jul 10;6:185.

[22] Kumar V, Varghese S. Ex Vivo Tumor-on-a-Chip Platforms to Study Intercellular Interactions within the Tumor Microenvironment. Adv Healthc Mater. 2019 Feb;8(4):e1801198.

[23] Hachey SJ, Hughes CCW. Applications of tumor chip technology. Lab Chip. 2018 Sep 26;18(19):2893-2912.

[24] Sung JH, Wang YI, Narasimhan Sriram N, Jackson M, Long C, Hickman JJ, Shuler ML. Recent Advances in Body-on-a-Chip Systems. Anal Chem. 2019 Jan 2;91(1):330-351.

[25] Skardal A, Shupe T, Atala A. Organoid-on-achip and body-on-a-chip systems for drug screening and disease modeling. Drug Discov Today. 2016 Sep;21(9):1399-1411.

[26] Boeri L, Izzo L, Sardelli L, Tunesi M, Albani D, Giordano C. Advanced Organ-on-a-Chip Devices to Investigate Liver Multi-Organ Communication: Focus on Gut, Microbiota and Brain. Bioengineering (Basel). 2019 Sep 28;6(4).

[27] https://ncats.nih.gov/tissuechip/projects/space (accessed 30.08.19).

[28] Tanaka K, Nishimura N, Kawai Y. Adaptation to microgravity, deconditioning, and countermeasures. J Physiol Sci. 2017 Mar;67(2):271-281.

[29] Fitzgerald J. Cartilage breakdown in microgravity-a problem for long-term spaceflight? NPJ Regen Med. 2017 Apr 11;2:10.

[30] Watenpaugh DE, Hargens AR (1996) The cardiovascular system in microgravity. In: Fregly MJ, Blatteis CM (eds) Handbook of physiology, the

gravitational environment. American Physiological Society, Maryland.

[31] Nicogossian, A.E., Williams, R.S., Huntoon, C.L., Doarn, C.R., Polk, J.D., Schneider, V.S. Space Physiology and Medicine. From Evidence to Practice. Springer-Verlag, New York, 2016.

[32] Mermel LA. Infection prevention and control during prolonged human space travel. Clin Infect Dis. 2013 Jan;56(1):123-30.

[33] Mukhopadhyay S, Saha R, Palanisamy A, Ghosh M, Biswas A, Roy S, Pal A, Sarkar K, Bagh S. A systems biology pipeline identifies new immune and disease related molecular signatures and networks in human cells during microgravity exposure. Sci Rep. 2016 May 17;6:25975.

[34] zayzafoon M, Meyers VE, McDonald JM. Microgravity: the immune response and

bone. Immunol Rev. 2005 Dec;208:267-80.

[35] Chang TT, Spurlock SM, Candelario TL, Grenon SM and Hughes-Fulford M. Spaceflight impairs antigenspecific tolerance induction in vivo and increases inflammatory cytokines. FASEB J. 2015;29:4122-32.

[36] Martinez EM, Yoshida MC, Candelario TL and Hughes-Fulford M. Spaceflight and simulated microgravity

cause a significant reduction of key gene expression in early T-cell activation. Am J Physiol Regul Integr Comp

Physiol. 2015;308:R480-8.

[37] Li M, Yao D, Zeng X, Kasakovski D, Zhang Y, Chen S, Zha X, Li Y, Xu L. Age related human T cell subset evolution and senescence. Immun Ageing. 2019 Sep 11;16:24.

[38] Fahy GM, Brooke RT, Watson JP, Good Z, Vasanawala SS, Maecker H, Leipold MD, Lin DTS, Kobor MS, Horvath S. Reversal of epigenetic aging and immunosenescent trends in humans. Aging Cell. 2019 Sep 8:e13028.

[39] Bucher CH, Schlundt C, Wulsten D, Sass FA, Wendler S, Ellinghaus A, Thiele T, Seemann R, Willie BM, Volk HD, Duda GN, Schmidt-Bleek K. Experience in the Adaptive Immunity Impacts Bone Homeostasis, Remodeling, and Healing. Front Immunol. 2019 Apr 12;10:797.

[40] Sattler S, Rosenthal N. The neonate versus adult mammalian immune system in cardiac repair and regeneration. Biochim Biophys Acta. 2016 Jul;1863(7 Pt B):1813-21.

[41] Scheuring RA, Mathers CH, Jones JA, Wear ML. Musculoskeletal injuries and minor trauma in space: incidence and injury mechanisms in U.S. astronauts. Aviat Space Environ Med. 2009;80(2):117-24. Wear M. Injury Rate of Shuttle Astronauts. The Longitudinal Study of Astronaut Health Newsletter. 1999;8(2):1-4.

[42] Willey JS, Kwok AT, Moore JE, et al. Spaceflight-Relevant Challenges of Radiation and/or Reduced Weight Bearing Cause Arthritic Responses in Knee Articular Cartilage. Radiat Res 2016;186:333-44.

[43] Keller TS, Strauss AM, Szpalski M. Prevention of bone loss and muscle atrophy during manned space flight. Microgravity Q 1992;2:89-102.

[44] Vandenburgh H, Chromiak J, Shansky J, et al. Space travel directly induces skeletal muscle atrophy. FASEB J 1999;13:1031-8.

[45] Punzi L, Galozzi P, Luisetto R, Favero M, Ramonda R, Oliviero F, Scanu A. Post-traumatic arthritis: overview on pathogenic mechanisms and role of inflammation. RMD Open. 2016 Sep 6;2(2):e000279.

[46] Liakopoulos V, Leivaditis K, Eleftheriadis T, Dombros N. The kidney in space. Int Urol Nephrol. 2012;44(6):1893-901.

[47] Cirillo M, De Santo NG, Heer M, Norsk P, Elmann-Larsen B, Bellini L, et al. Urinary albumin in space missions. J Gravit Physiol. 2002;9(1):P193-4.

[48] Drummer C, Cirillo M, De Santo NG. History of fluid balance and kidney function in space. J Nephrol. 2004;17(1):180-6.

[49] Breyer MD, Susztak K. The next generation of therapeutics for chronic kidney disease. Nat Rev Drug Discov. 2016 Aug;15(8):568-88.

[50] Yeung CK, Koenig P, Countryman S, Thummel KE, Himmelfarb J, Kelly EJ. Tissue Chips in Space-Challenges and Opportunities. Clin Transl Sci. 2019 Sep 16.

[51] Vogel J, Thiel CS, Tauber S, Stockmann C, Gassmann M, Ullrich O. Expression of Hypoxia-Inducible Factor 1α (HIF- 1α) and Genes of Related Pathways in Altered Gravity. Int J Mol Sci. 2019 Jan 20;20(2). pii: E436.

[52] Bellone JA, Gifford PS, Nishiyama NC, Hartman RE, Mao XW. Long-term effects of simulated microgravity and/or chronic exposure to low-dose gamma radiation on behavior and blood-brain barrier integrity. NPJ Microgravity. 2016 Jun 9;2:16019.

[53] Rivera FJ, Hinrichsen B, Silva ME. Pericytes in Multiple Sclerosis. Adv Exp Med Biol. 2019;1147:167-187.

[54] Marchi N, Granata T, Ghosh C, Janigro D. Blood-brain barrier dysfunction and epilepsy: pathophysiologic role and therapeutic approaches. Epilepsia. 2012 Nov;53(11):1877-86.

[55] Sonnenfeld G, Mandel AD, Konstantinova IV, Berry WD, Taylor GR, Lesnyak AT, Fuchs BB, Rakhmilevich AL. Spaceflight alters immune cell function and distribution. J Appl Physiol (1985). 1992 Aug;73(2 Suppl):191S-195S.

[56] Sonnenfeld G, Davis S, Taylor GR, Mandel AD, Konstantinova IV, Lesnyak A, Fuchs BB, Peres C, Tkackzuk J, Schmitt DA. Effect of space flight on cytokine production and other immunologic parameters

of rhesus monkeys. J Interferon Cytokine Res. 1996 May;16(5):409-15.

[57] Peiró T, Patel DF, Akthar S, Gregory LG, Pyle CJ, Harker JA, Birrell MA, Lloyd CM, Snelgrove RJ. Neutrophils drive alveolar macrophage IL-1 β release during respiratory viral infection. Thorax. 2018 Jun;73(6):546-556.

[58] Fitts RH, Trappe SW, Costill DL, Gallagher PM, Creer AC, Colloton PA, Peters JR, Romatowski JG, Bain JL, Riley DA. Prolonged space flight-induced alterations in the structure and function of human skeletal muscle fibres. J Physiol. 2010 Sep 15;588(Pt 18):3567-92.

[59] Trappe S, Costill D, Gallagher P, Creer A, Peters JR, Evans H, Riley DA, Fitts RH. Exercise in space: human skeletal muscle after 6 months aboard the International Space Station. J Appl Physiol (1985). 2009 Apr;106(4):1159-68.

[60] Vandenburgh H, Chromiak J, Shansky J, Del Tatto M, Lemaire J. Space travel directly induces skeletal muscle atrophy. FASEB J. 1999 Jun;13(9):1031-8.

[61] Larsson L, Degens H, Li M, Salviati L, Lee YI, Thompson W, Kirkland JL, Sandri M. Sarcopenia: Aging-Related Loss of Muscle Mass and Function. Physiol Rev. 2019 Jan 1;99(1):427-511.

[62] Aversa Z, Zhang X, Fielding RA, Lanza I, LeBrasseur NK. The clinical impact and biological mechanisms of skeletal muscle aging. Bone. 2019 Oct;127:26-36.

[63] Otsuka K, Cornelissen G, Kubo Y, Hayashi M, Yamamoto N, Shibata K, Aiba T, Furukawa S, Ohshima H, Mukai C. Intrinsic cardiovascular autonomic regulatory system of astronauts exposed long-term to microgravity in space: observational study. NPJ Microgravity. 2015 Nov 30;1:15018.

[64] Otsuka K, Cornelissen G, Furukawa S, Kubo Y, Hayashi M, Shibata K, Mizuno K, Aiba T, Ohshima H, Mukai C. Long-term exposure to space's microgravity alters the time structure of heart rate variability of astronauts. Heliyon. 2016 Dec 19;2(12):e00211.

[65] Hughson RL, Robertson AD, Arbeille P, Shoemaker JK, Rush JW, Fraser KS, Greaves DK. Increased postflight carotid artery stiffness and inflight insulin resistance resulting from 6-mo spaceflight in male and female astronauts. Am J Physiol Heart Circ Physiol. 2016 Mar 1;310(5):H628-38.

[66] Gude NA, Sussman MA. Cardiac regenerative therapy: Many paths to repair. Trends Cardiovasc Med. 2019 Sep 2. pii: S1050-1738(19)30127-6.

[67] Benjamin EJ, Blaha MJ, Chiuve SE, Cushman M, Das SR, Deo R, de Ferranti SD, Floyd J, Fornage M, Gillespie C, Isasi CR, Jiménez MC, Jordan LC, Judd SE, Lackland D, Lichtman JH, Lisabeth L, Liu S, Longenecker CT, Mackey RH, Matsushita K, Mozaffarian D, Mussolino ME, Nasir K, Neumar RW,

Palaniappan L, Pandey DK, Thiagarajan RR, Reeves MJ, Ritchey M, Rodriguez CJ, Roth GA, Rosamond WD, Sasson C, Towfighi A, Tsao CW, Turner MB, Virani SS, Voeks JH, Willey JZ, Wilkins JT, Wu JH, Alger HM, Wong SS, Muntner P; American Heart Association Statistics Committee and Stroke Statistics Subcommittee. Heart Disease and Stroke Statistics-2017 Update: A Report From the American Heart Association. Circulation. 2017 Mar 7;135(10):e146-e603.

[68] Sides MB, Vernikos J, Convertino VA, Stepanek J, Tripp LD, Draeger J, Hargens AR, Kourtidou-Papadeli C, Pavy-LeTraon A, Russomano T, Wong JY, Buccello RR, Lee PH, Nangalia V, Saary MJ. The Bellagio Report: Cardiovascular risks of spaceflight: implications for the future of space travel. Aviat Space Environ Med. 2005 Sep;76(9):877-95.

[69] Diakos NA, Selzman CH, Sachse FB, Stehlik J, Kfoury AG, Wever-Pinzon O, Catino A, Alharethi R, Reid BB, Miller DV, Salama M, Zaitsev AV, Shibayama J, Li H, Fang JC, Li DY, Drakos SG. Myocardial atrophy and chronic mechanical unloading of the failing human heart: implications for cardiac assist device-induced myocardial

recovery. J Am Coll Cardiol. 2014 Oct 14;64(15):1602-12.

[70] Jordaens L. A clinical approach to arrhythmias revisited in 2018 : From ECG over noninvasive and invasive electrophysiology to advanced imaging. Neth Heart J. 2018;26(4):182–189.

[71] Sawyer HR, Moeller CL, Phillips RW, Smirnov KL. Proliferation of jejunal mucosal cells in rats flown in space. J Appl Physiol (1985). 1992 Aug;73(2 Suppl):148S-150S.

[72] Garrett-Bakelman FE, Darshi M, Green SJ, Gur RC, Lin L, Macias BR, McKenna MJ, Meydan C, Mishra T, Nasrini J, Piening BD, Rizzardi LF, Sharma K, Siamwala JH, Taylor L, Vitaterna MH, Afkarian M, Afshinnekoo E, Ahadi S, Ambati A, Arya M, Bezdan D, Callahan CM, Chen S, Choi AMK, Chlipala GE, Contrepois K, Covington M, Crucian BE, De Vivo I, Dinges DF, Ebert DJ, Feinberg JI, Gandara JA, George KA, Goutsias J, Grills GS, Hargens AR, Heer M, Hillary RP, Hoofnagle AN, Hook VYH, Jenkinson G, Jiang P, Keshavarzian A, Laurie SS, Lee-McMullen B, Lumpkins SB, MacKay M, Maienschein-Cline MG, Melnick AM, Moore TM, Nakahira K, Patel HH, Pietrzyk R, Rao V, Saito R, Salins DN, Schilling JM, Sears DD, Sheridan CK, Stenger MB, Tryggvadottir R, Urban AE, Vaisar T, Van Espen B, Zhang J, Ziegler MG, Zwart SR, Charles JB, Kundrot CE, Scott GBI, Bailey SM, Basner M, Feinberg AP, Lee SMC, Mason CE, Mignot E, Rana BK, Smith SM, Snyder MP, Turek FW. The NASA Twins Study: A multidimensional analysis of a year-long human spaceflight. Science. 2019 Apr 12;364(6436). pii: eaau8650.

[73] Bagga D, Reichert JL, Koschutnig K, Aigner CS, Holzer P, Koskinen K, Moissl-Eichinger C, Schöpf V. Probiotics drive gut microbiome triggering emotional brain signatures. Gut Microbes. 2018 Nov 2;9(6):486-496.

[74] Amedei A, Barceló-Coblijn G. Editorial of Special Issue "The Interplay of Microbiome and

Immune Response in Health and Diseases". Int J Mol Sci. 2019 Jul 29;20(15). pii: E3708.