

Marc Giulianotti¹, Amelia W. Smith, and Debbie Wells Center for the Advancement of Science in Space ¹To whom correspondence should be addressed. Email: mgiulianotti@iss-casis.org

OCTOBER 24, 2017

TECHNICAL CORRESPONDENCE FROM THE INTERNATIONAL SPACE STATION U.S. NATIONAL LABORATORY

Microgravity Molecular Crystal Growth Onboard the ISS National Lab: **A Program Overview**

EXECUTIVE SUMMARY

The International Space Station (ISS) U.S. National Laboratory provides a valuable platform for improved molecular crystal growth, and the many successful crystallization experiments conducted in space over the past three decades have demonstrated this value. Crystals grown in microgravity are often larger and more wellordered than Earth-grown crystals.

In the analysis of organic molecules, high-quality crystals can lead to improved datasets for structure resolution, mosaicity (misalignment within the crystal lattice), and electron density. Larger crystals can also enable analysis by neutron diffraction (instead of more traditional diffraction analyses, which use X-rays), which provides greater structural detail by allowing for the determination of hydrogen positions within protein structures. For inorganic molecules, high-quality crystals can lead to advances in metal manufacturing, electronics, and radiation detection.

Several commercial entities, including pharmaceutical companies Merck & Co. and Eli Lilly and Company, are currently using the unique crystallization environment onboard the ISS National Lab to advance their research and development (R&D). The Center for the Advancement of Science in Space (CASIS) is committed to establishing an ISS National Lab Microgravity Molecular Crystal Growth (MMCG) Program to enable continued use of the ISS for crystallization studies and to enhance the commercialization potential of low Earth orbit (LEO) platforms for both organic and inorganic molecular crystallization.

To begin to outline the requirements and lay the foundation for a sustainable MMCG Program, CASIS held a technical interchange meeting in 2015 to gather input from experts in the field of protein crystallography. Combining lessons learned from past spaceflight crystallization R&D with expert recommendations from the interchange meeting, CASIS established criteria for informed molecule selection to aid in the identification of optimal organic molecules to study in microgravity (see the section titled "Expert Input and Informed Molecule Selection" for additional information). While these initial criteria apply only to organic molecule crystallization, CASIS also intends to gather expert input from the inorganic crystallization community to optimize the success of space-based inorganic crystal growth research that may benefit life on Earth.

The ISS National Lab MMCG Program specifically aims to provide the following:

- Opportunities for molecular crystal growth investigations on every cargo resupply (CRS) launch to the ISS.
- Rapid turnaround of samples—a 90-day cycle time from molecule identification to crystal return from the ISS.
- Hardware options that minimize preflight optimization steps through use of standard laboratory crystallization procedures to enhance readiness for analyses.
- Processes to support multi-year customer programs for crystallization.

The sections that follow provide a brief history of crystallization in microgravity, an overview of crystal growth investigations conducted onboard the ISS National Lab, and additional information on the CASIS technical interchange meeting and resulting expert recommendations. Also provided is a summary of applications for molecular crystal growth in microgravity, additional information on the MMCG Program and its goals and implementation, an overview of continued interest in microgravity molecular crystal growth research, and a discussion of future directions.

BACKGROUND

Microgravity has been used for more than 30 years to improve outcomes of molecular crystal growth—on U.S. space shuttle missions, onboard the Russian space station Mir, on free-flying spacecraft, and onboard the ISS.

Organic Macromolecular Crystal Growth in Microgravity

Observed benefits of organic crystallization in microgravity include larger, more well-ordered crystals with a higher diffraction resolution and lower mosaicity. The electron density maps from microgravity-grown crystals are often more detailed than those from Earth-grown crystals, improving the accuracy of protein structure determination. Researchers hypothesize that these improvements are due to the slower, more ordered diffusion-driven movement of molecules during crystallization in microgravity resulting in a more uniform incorporation of molecules into the crystalline lattice.

The first organic crystallization experiments in microgravity were conducted on U.S. space shuttle missions in the mid-1980s. In these early experiments, results were not definitive, and some crystals benefitted from growth in microgravity while others did not. However, these experiments demonstrated the central role that gravity plays in crystal growth and spurred interest within the crystallography community in the potential for microgravity to improve macromolecular crystal growth. Several factors limited these crystallization experiments, including the short duration and infrequency of the shuttle missions, uncertainty in the launch schedule, limited sample capacity and experiment volume, and instability in temperature. Conducting crystallization experiments onboard the ISS held promise to address several of these issues.

Crystallization experiments on the shuttle and Mir in the 1990s and onboard the ISS in the early 2000s further validated microgravity as a key factor in improved crystal growth in space. These experiments yielded numerous large-volume crystals with higher diffraction resolution and vast improvement in mosaicity. However, results continue to be mixed, with only some proteins exhibiting improved growth in microgravity.

Inorganic Crystallization in Microgravity

The elimination of gravity-driven forces (e.g., convection, sedimentation, and buoyancy) in microgravity also results in the synthesis of inorganic crystals that are larger and more well-ordered than crystals synthesized on the ground. Experiments conducted on sounding rockets and the shuttle in the 1990s, and later on the ISS, highlighted gravity as a key source of disturbing effects that occur during ground-based alloy solidification. These studies provided

valuable benchmark data for validating solidification models. Experiments performed on the shuttle in the 1990s also sought to elucidate the effects of gravity on colloidal crystallization. These studies confirmed that gravity alters inherent aspects of colloidal crystallization, and investigators were able to crystallize samples that had not yet been crystallized on the ground in a matter of days in microgravity. Additionally, as with organic molecules, crystallization in microgravity yielded larger, higherquality crystals, some exhibiting dendritic arms previously undetected in crystallization on the ground.

Additional solidification experiments conducted on the ISS in the 2000s found that adding magnetic fields to solidification in microgravity results in high-performance magnetostrictive materials. Other experiments used the Solidification Using a Baffle in Sealed Ampoules (SUBSA) furnace on the ISS to synthesize semiconductor crystals, facilitating the observation of melting and directional solidification in microgravity—toward ultimate application in electronic devices such as computers, medical imagers, and radiation detectors.

Crystallization Investigations on the ISS National Lab

The ISS National Lab has continued to provide a valuable platform for both organic and inorganic crystal growth research in microgravity and has enabled many crystallization successes to date.

Initial CASIS-sponsored organic crystal growth investigations conducted on the ISS National Lab stemmed from a 2012 CASIS solicitation for nextgeneration space-based protein crystallization research, and the resulting investigations were launched to the ISS National Lab on SpaceX CRS-3 and CRS-4 in 2014. Two of these investigations produced protein crystals of sufficient size for neutron diffraction, and many produced high-quality crystals for analysis. Since those initial flights, many additional commercial and academic crystal growth investigations have flown to the ISS National Lab (see Table 1). The majority of these investigations were focused on structural determination for functional studies and structure-based drug design, but others, including multiple investigations from the pharmaceutical company Merck, are using the improved crystal growth onboard the ISS National Lab for studies aimed at improving drug formulation, manufacturing, and storage. In the case of pharmaceutical researchers, multiple flight opportunities to the ISS National Lab in rapid succession provides an opportunity for investigators to adjust experimental design and maximize success of their R&D objectives by



iteration. Moreover, a decrease in the time from proposal submission to flight allows companies with fast-paced R&D goals to obtain relevant results in a timely manner. For example, a recent project from the Michael J. Fox Foundation flew to the ISS National Lab less than one year after initial proposal submission—in fact, from specific molecule identification to flight, the process took less than three months.

Additionally, in 2016 a significant milestone in the effort to validate organic crystallization systems for microgravity that use common laboratory hardware was achieved as part of a CASIS-sponsored investigation by the pharmaceutical company Eli Lilly and Company. This investigation, which launched to the ISS National Lab on SpaceX CRS-8 in 2016, marked the first flight of a commercial-off-the-shelf (COTS) protein crystallization plate, developed by the biotechnology company MiTeGen. Expanding the use of COTS hardware is an important step toward streamlining preflight optimization work—lowering costs and shortening the time to crystal retrieval.

With respect to inorganic crystallization experiments launched to the ISS National Lab, two notable investigations utilizing the recently refurbished SUBSA furnace launched on Orbital ATK-7 in 2016. One of these investigations, led by Dr. Aleksandar Ostrogorsky of the Illinois Institute of Technology (who conducted some of the original ISS SUBSA experiments in 2002), sought to produce new types of semiconductor crystals, and the other, led by Dr. Alexei Churilov of Radiation Monitoring Devices, Inc., aimed to synthesize high-quality scintillation crystals—both seeking to improve materials for use in advanced radiation detection devices for homeland security applications.

TABLE 1: ISS NATIONAL LAB CRYSTALLIZATION INVESTIGATIONS

Investigation*	Launch Vehicle/Date	Aim	
TITLE: Protein Crystals for Neutron Crystallography: Large Volume Crystal Growth for Inorganic Pyrophosphatase Complexes by Counter-Diffusion in Microgravity for Neutron Diffraction Studies (PC4NC)	SpX-3, 4/18/2014	Produce crystals of inorganic pyrophosphate phosphatase (IPPase) of sufficient size for neutron diffraction, toward structural determination of this medically important protein	
PRINCIPAL INVESTIGATOR: Joseph Ng; iXpressGenes, Inc.			
TITLE: Microgravity Growth of Single Crystals for Structure Determination (Merck PCG)	SpX-3, 4/18/2014	Produce crystals of a human monoclonal antibody undergoing clinical trials for treatment of immunological disease, toward improved structural	
PRINCIPAL INVESTIGATOR: Paul Reichert; Merck Research Laboratories		determination	
TITLE: Crystallization of Human Membrane ABC Proteins in Microgravity (CASIS PCG HDPCG-2)	SpX-3, 4/18/2014	Produce crystals of medically important membrane proteins, including cystic fibrosis protein, toward	
PRINCIPAL INVESTIGATOR: Stephen Aller; University of Alabama at Birmingham		structural determination	
TITLE: Crystallization of Huntingtin Exon-1 Using Microgravity (CASIS PCG HDPCG-1)	SpX-3, 4/18/2014	Produce crystals of huntingtin, a protein associated with Huntington's Disease, toward structural determination of regions that are difficult to crystallize on the ground	
PRINCIPAL INVESTIGATOR: Pamela Bjorkman; University of Alabama at Birmingham			
TITLE: Crystallization of Medically Relevant Proteins Using Microgravity (CASIS PCG GCF-2a)	SpX-3, 4/18/2014	Produce improved crystals of two medically important proteins, toward structural determination	
PRINCIPAL INVESTIGATOR: Sergey Korolev; St. Louis University			
TITLE: Exploiting on-orbit crystal properties for structural studies of medically and economically important targets (CASIS PCG GCF-2b)	SpX-3, 4/18/2014	Produce crystals of four medically important proteins, toward structural determination	
PRINCIPAL INVESTIGATOR: Edward Snell; Hauptman Woodward Medical Research Institute, Inc.			
TITLE: Optimization of Protein Crystal Growth for Determination of Enzyme Mechanisms through Advanced Diffraction Techniques (CASIS PCG 2-1)	SpX-4, 9/21/2014	Produce crystals of three medically important proteins of sufficient size for neutron diffraction, toward structural determination	
PRINCIPAL INVESTIGATOR: Constance Schall; University of Toledo			
TITLE: A collaborative proposal for protein crystal growth in space to enable therapeutic discovery (Module-19 S/N 1002)	SpX-4, 9/21/2014	Produce crystals of two challenging therapeutic targets implicated in cardiovascular disease and cancer, toward structural determination	
PRINCIPAL INVESTIGATORS : Cory Gerdts; Protein BioSolutions and Matt Clifton; Beryllium Discovery Corp.			

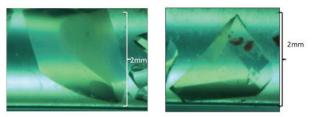
*Operational Nomenclature in parentheses.



Investigation*	Launch Vehicle/Date	Aim	
TITLE: Microgravity Growth of Crystalline Monoclonal Antibodies for Pharmaceutical Applications (CASIS PCG 3)	SpX-6, 6/28/2015	Produce crystals of medically important human monoclonal antibodies, toward improved drug formulation, manufacturing, and storage	
PRINCIPAL INVESTIGATOR: Paul Reichert; Merck Research Laboratories			
TITLE: A Co-Crystallization in Microgravity Approach to Structure- Based Drug Design (CASIS PCG 4-1)	SpX-8, 4/8/2016	Produce high-quality crystals of a medically important protein in complex with a small molecule ligand (potential therapeutic), toward improved structure-based drug design	
PRINCIPAL INVESTIGATOR: Kristopher Gonzalez-DeWhitt; Eli Lilly and Company			
TITLE: The Effect of Microgravity on the Co-Crystallization of a Membrane Protein with a Medically Relevant Compound (CASIS PCG 4-2)	SpX-8, 4/8/2016	Crystallize protein complexes consisting of a medically important membrane protein bound to a potential therapeutic compound, toward improved	
PRINCIPAL INVESTIGATOR: Michael Hickey; Eli Lilly and Company		structure-based drug design	
TITLE: The Effect of Microgravity on the Electrolysis of Silver Nitrate NR Module-69)	SpX-9, 7/18/2016	Analyze the three-dimensional structure of silver crystals in microgravity to determine whether elimination of gravity produces higher-quality	
PRINCIPAL INVESTIGATOR: David Schlichting; Eaglecrest High School		crystals (student investigation)	
IITLE: Detached Melt and Vapor Growth of InI in SUBSA Hardware IIT – SUBSA)	Orbital ATK-7, 9/15/2016	Synthesize new types of semiconductor crystals, toward advanced radiation detection capabilities	
PRINCIPAL INVESTIGATOR: Aleksandar Ostrogorsky; Illinois Institute of Technology			
TITLE: Crystal Growth of Cs2LiYCl6:Ce in Microgravity (RMD – SUBSA)	Orbital ATK-7, 9/15/2016	Produce high-quality scintillation crystals, toward advanced radiation detection capabilities	
PRINCIPAL INVESTIGATOR: Alexei Churilov; Radiation Monitoring Devices, Inc.			
TITLE: Microgravity Growth of Crystalline Monoclonal Antibodies for Pharmaceutical Applications (CASIS PCG 5)	SpX-10, 2/19/2017	Produce high-quality, uniform crystalline suspensions of a monoclonal antibody PD-1 drug, Keytruda, toward improved drug formulation, manufacturing, and storage	
PRINCIPAL INVESTIGATOR: Paul Reichert; Merck Research Laboratories			
TITLE: The Effect of Macromolecular Transport on Microgravity Protein Crystallization (LMM Biophysics 1)	SpX-10, 2/19/2017	Validate the hypothesis that the improved quality of microgravity-grown biological crystals is the result of two macromolecular characteristics that exist in buoyancy-free, diffusion-dominated solution	
PRINCIPAL INVESTIGATOR: Lawrence DeLucas; University of Alabama at Birmingham			
TITLE: Growth Rate Dispersion as a Predictive Indicator for Biological Crystal Samples Where Quality can be Improved with Microgravity Growth (LMM Biophysics 3)	SpX-10, 2/19/2017	Validate the hypothesis that growth rate dispersion could be an indicator of crystals whose quality may be improved in microgravity	
PRINCIPAL INVESTIGATOR: Edward Snell; Hauptman Woodward Medical Research Institute, Inc.			
TTLE: Neutron Crystallographic Studies of Human Acetylcholinesterase for the Design of Accelerated Reactivators CASIS PCG 6)	SpX-11, 6/3/2017	Produce crystals of a medically important protein of sufficient quality for neutron diffraction, toward structural determination	
PRINCIPAL INVESTIGATOR: Andrey Kovalevsky; Dak Ridge National Laboratory			
TITLE: Crystallization of LRRK2 under Microgravity Conditions CASIS PCG 7)	SpX-12, 8/14/2017	Crystallize LRRK2, a protein associated with Parkinson's disease, toward structural determination of this difficult-to-crystallize protein	
PRINCIPAL INVESTIGATOR: Marco Baptista; Michael J. Fox Foundation			

*Operational Nomenclature in parentheses.





Joseph Ng of iXpressGenes, Inc. flew crystals of IPPase on SpaceX CRS-3, obtaining large crystals suitable for neutron diffraction.



Expert Input and Informed Molecule Selection

To continue to support the growing demand for crystallization onboard the ISS National Lab and begin to outline the basic requirements for a long-term crystallization program in space, CASIS held a technical interchange meeting in 2015 to gather in-depth feedback from experts across the field of protein crystallography. The technical interchange meeting focused on accessibility and timing, flight and ground resources, parallel education initiatives, and funding. Attendees discussed past and current crystallization research efforts, current technologies and capabilities in the field, and the needs of the protein crystallography community on the ISS National Lab. Information gained from the technical interchange initiated the path forward to implementation of an ISS National Lab MMCG Program to enable sustainable, repetitive, low-cost crystallization in microgravity. Details from this event can be found online (http://www.spacestationresearch.com/ research-library/reports/2015pcg/).

Technical interchange invitees included former and current microgravity crystallographers, ground-based crystallographers, crystallography analytics experts, and crystallography hardware vendors. Key discussion points and recommendations from the technical interchange that helped to lay the foundation for the development of the MMCG Program and drive the program's goals are listed in Box 1, and a process for informed molecule selection for organic crystallization in microgravity was formulated based on input from the technical interchange.

Informed molecule selection criteria for the ISS National Lab MMCG Program:

- 1. The protein should be able to achieve nucleation on the ground, preferably in the flight hardware.
- 2. The protein should be able to be produced in enough quantity and with enough uniformity to support experiment/operational design and improve the chances for successful crystallization. Material for at least three scrub refurbishments should be planned, and contingency plans should be in place in the event of additional delays.
- **3**. If the user has a crystal of the protein, there should be a demonstrated need for improvement (e.g., larger or more uniformly produced crystals and improved diffraction resolution or electron density map). Examples include crystals that have a current resolution less than 4.0 angstroms or current crystals that diffract at 2.5 angstroms but higher quality is needed to provide additional data.
- The user should know some characteristics of the protein's stability, such as temperature range and timedependent deterioration.

BOX 1: RECOMMENDATIONS FROM THE TECHNICAL INTERCHANGE MEETING

- The comparative advantage of crystal growth in microgravity should be documented, and outreach to the crystallography community stating these advantages (backed up by literature) is critical.
- Molecules of interest include any protein of high biological significance with an indication of scientific, medical, or commercial interest with a need for improved diffraction and/or electron density map. Molecules of interest include both organic molecules (e.g., membrane proteins, protein-protein and multi-protein complexes, protein-ligand interactions, enzymes, ion channel proteins, side chain configuration of proteins, and small molecules) and inorganic molecules (e.g., semiconductor/liquid crystals and zeolites).
- There is a need to grow both large crystals and small uniform crystals to meet a variety of research and manufacturing objectives, thus hardware and processes must be available to support both.
- It is crucial to provide reliable in-orbit access and secure return of samples to the user's lab or analyses location. It is also important to provide repetitive access for each user; typically, multiple flights are required to achieve crystals resulting in increased resolution of structure.
- There should be a focus on down-selecting to a small number of hardware pieces that work best and ensuring they are fully enabled to fly experiments rapidly.
- Although imaging is not required, it would be useful for both the science (e.g., knowing when nucleation took place or when the crystal started to degrade after growth) as well as educational outreach.
- A related science, technology, engineering, and mathematics (STEM) education program should expand reach and engage new funding opportunities.
- Support is needed for several crystallization techniques, including vapor diffusion, liquidliquid diffusion, and batch crystallization for X-ray diffraction and neutron diffraction (the key analytic tools used by crystallographers).
- It is essential to reduce the "learning curve" for new users to translate laboratory-based crystallization successes to flight configurations. Efforts should be made to make crystallization compatible with the industry standard Society for Laboratory Automation and Screening (SLAS), formerly the Society for Biomolecular Screening (SBS), plates and automation.

THE ISS NATIONAL LAB MICROGRAVITY MOLECULAR CRYSTAL GROWTH PROGRAM

Progress has been made in the development and implementation of the ISS National Lab MMCG Program, which will provide a platform for discovery to users across many communities—commercial, government, academia, and private research—while also supporting future efforts toward the commercialization of LEO.

Applications for Molecular Crystal Growth in Microgravity

The MMCG Program encompasses both organic and inorganic crystallization, each with numerous applications (see Table 2). Organic microgravity crystallization has applications in the pharmaceutical and agricultural industries. The majority of pharmaceutical applications are aimed at protein structure determination for improved drug development. Many crystals grown in microgravity are larger and more detailed than those grown on Earth, providing protein structures that are more accurate, thus enabling better structure-based drug designs. Other pharmaceutical applications seek to produce highquality uniform crystalline suspensions for improved drug formulation, manufacturing, and storage. Agricultural applications are aimed at developing improved agriculture formulations for pest control and growth enhancement.

Inorganic crystallization in microgravity also has a multitude of applications. Microgravity enables the synthesis of high-quality semiconductor and scintillator crystals, which can be used for improved radiation detection, with applications in homeland security and medical imaging devices. Synthesis of higher-quality semiconductor crystals also leads to advances in electronics such as computers and smart phones. Alloy solidification research in microgravity is aimed at understanding how defects form during the solidification process and can lead to advances in metal manufacturing. Microgravity studies of electrochemical deposition (the process by which an electric current is used to form thin metal features on conductive surfaces such as electrodes) can help to elucidate the role gravity plays in the formation of imperfections on conductive surfaces, toward production of high-aspect-ratio structures with fewer imperfections.

Goals of the ISS National Lab MMCG Program

The overarching goal of the ISS National Lab MMCG Program is to implement processes to increase commercial utilization of LEO in the area of molecular crystal growth. The program seeks to make it easier and more economical for commercial users to take advantage of microgravity to advance their R&D objectives.

TABLE 2: APPLICATIONS FOR MICROGRAVITY CRYSTALLIZATION

ORGANIC		Structural determination for drug development	
	Pharmaceutical	Uniform crystalline suspensions for drug formulation and delivery, manufacturing, and storage	
	Agricultural	Agriculture formulation development	
Radiation		Semiconductor crystal	
		growth	
	Radiation detection	growth Scintillator crystal growth	
INORGANIC			

For organic molecular crystallization, the MMCG Program was established with the following goals:

- To provide repeat flight opportunities to investigators who fit a predefined set of criteria indicating that microgravity may help. This criteria for "informed molecule selection" was set forth by experts in the field of crystallography who attended the CASIS protein crystal growth technical interchange meeting. The goal of the MMCG Program is not to replace crystallization techniques currently used on the ground, but rather to inform crystallographers of the ideal conditions in which microgravity could be a valuable tool to improve crystallization.
- To streamline access to the ISS National Lab through dedicated launch opportunities, rapid turnaround of samples, and cost-effective set pricing. The MMCG Program aims to fly and return samples within a three-month timeframe from the time of molecule construct determination, enabling a rapid return of samples to the research pathway.
- To minimize risk and increase the probability of success through expanded use and validation of COTS hardware and through access to vetted service providers with expertise in microgravity investigation design. The use of COTS hardware reduces both the cost of flight hardware as well as time and money spent on optimizing conditions.
- To work toward multi-flight, multi-protein collaborations with other government organizations, research foundations, and commercial companies. The MMCG Program has been specifically designed to meet the needs of commercial upstream R&D, and as a result, CASIS is already beginning to negotiate several multi-year, multiflight programs.



A first step in developing the MMCG Program was to engage existing and potential new partners intent on providing relevant hardware and services. Toward this end, CASIS issued a solicitation in 2016 for proposals to provide support services—including laboratory services, integration, and hardware support—for investigators interested in conducting organic crystallization experiments onboard the ISS National Lab. The goals of the solicitation were to (1) standardize hardware for vapor diffusion, liquid-liquid diffusion, and batch crystallization; (2) identify service providers for specialized services such as protein expansion and crystal analyses; and (3) establish consistent fee-for-service with providers to streamline the customer experience.

As a result of the solicitation, the following preferred partners were chosen, and costs for hardware and implementation services were defined, locking prices for standard services for five years.

- ► The University of Alabama at Birmingham
- Teledyne Brown Engineering
- The Bionetics Corporation
- Techshot, Inc.

Likewise, CASIS intends to bring together the inorganic molecular crystallization research community to identify goals and needs to enhance the use of microgravity for inorganic crystal growth.

CUSTOMER ENGAGEMENT AND FUTURE DIRECTIONS

Increased outreach to the crystallography community will expand understanding of microgravity as a tool to improve molecular crystallization. To connect with the crystallography community, communicate the advantages of crystallization in microgravity, and provide information on the ISS National Lab MMCG Program, CASIS presented and exhibited at the 2016 American Crystallographic Association annual meeting—one of the largest gatherings of crystallographers-and exhibited at the 2017 annual meeting. CASIS also presented at the 2016 Frontiers in Structural Biology of Membrane Protein & Pittsburgh Diffraction Conference. Continued participation in key crystallography events is planned to further enable customer engagement and spur new interest in microgravity molecular crystal growth research (see Table 3). In addition, traditional targeted outreach to key thought leaders and commercial sectors is ongoing.

As a result of increased customer engagement, the MMCG Program continues to attract preeminent industry and academic researchers, as can be seen in the upcoming

TABLE 3: SELECT UPCOMING MMCG EVENTS

Event	Type of Engagement	Date	Location
American Society for Gravitational and Space Research (ASGSR) 2017 Annual Meeting	Symposium	10/28/17	Seattle, WA
American Crystallographic Association (ACA) 2018 Annual Meeting	Exhibit and panel session	7/22/18	Toronto, Canada
ISS Research and Development (R&D) 2018 Conference	Crystal growth technical sessions	7/23/18 – 7/26/18	San Francisco, CA

crystallization investigations planned for launch to the ISS National Lab within the next few increments (see Table 4). The multitude of commercial and academic investigators currently utilizing the microgravity environment on the ISS National Lab to advance their crystallization research demonstrates the effectiveness of CASIS outreach and the growing interest within the crystallography community. CASIS is committed to establishing an MMCG Program to support these investigators and enhance the commercialization potential of a LEO platform for crystallization research.

In summary, the MMCG Program aims to streamline access to the ISS National Lab by providing a reliable flight schedule, optimized hardware, rapid turnaround of samples, cost-effective set pricing, and repeat flight opportunities. CASIS is working with COTS hardware manufacturers and the user community to optimize current hardware designs to enable a smoother transition for users going from standard ground-based crystallization to spaceflight crystallization and to enhance readiness of samples for analyses. Moreover, to aid in the identification of optimal organic molecules for growth in microgravity, CASIS has developed criteria for informed molecule selection based on lessons learned from past spaceflight R&D and expert feedback—and additional investigations are underway to help further refine the process for informed molecule selection. Finally, CASIS is working toward establishing multi-flight collaborations with commercial entities, government agencies, and research foundations and has specifically designed the MMCG Program to meet the needs of commercial upstream R&D. As a result, CASIS is currently in the process of negotiating several multi-year, multi-flight programs.



TABLE 4: SELECT UPCOMING ISS NATIONAL LAB CRYSTALLIZATION INVESTIGATIONS

Investigation	Aim
TITLE: Nemak Alloy Solidification Experiments PRINCIPAL INVESTIGATOR: Dr. Glenn Byczynski; Nemak	Elucidate critical parameters behind the formation of hot tearing, which causes defects in the solidification of metal alloys, toward improvements in metal manufacturing
TITLE: 2017 Wisconsin Crystal Growing Competition and Lecture tour PRINCIPAL INVESTIGATOR: Ilia Guzei; University of Wisconsin, Madison	Provide a flight opportunity for the middle and high school student winners of the 2017 Wisconsin Crystal Growing Competition, allowing the students to test their optimized conditions for Earth-based crystallization against microgravity-based crystallization
TITLE: Microgravity Crystallization of Glycogen Synthase-Glycogenin Protein Complex PRINCIPAL INVESTIGATOR: Dr. David Chung; Dover Lifesciences	Crystallize a medically important protein in complex with inhibitor molecules, toward structural determination
TITLE: An ISS Experiment on Electrodeposition PRINCIPAL INVESTIGATOR: Dr. Kirk Ziegler; University of Florida	Elucidate the role of gravity in the formation of interfacial instability patterns during electrochemical deposition, the process by which an electric current is used to form thin metal features on conductive surfaces such as electrodes, toward production of high-aspect-ratio structures with fewer imperfections
TITLE: Microgravity Crystal Growth for Improvement in Neutron Diffraction PRINCIPAL INVESTIGATOR: Dr. Timothy Mueser; University of Toledo	Crystalize three medically important proteins with sufficient quality for neutron diffraction, toward structural determination
TITLE: Microgravity Investigation of Cement Solidification PRINCIPAL INVESTIGATOR: Dr. Aleksandra Radlinska; Penn State University	Study the microstructural development of cement, which occurs in stages during the hydration reaction and hardening process and results in elaborate combinations of different crystals, toward better understanding of the complex process of cement solidification

REFERENCES

- Abd Rahman RN, Ali MS, Sugiyama S, Leow AT, Inoue T, Basri M, Salleh AB, Matsumura H. A comparative analysis of microgravity and earth grown thermostable T1 lipase crystals using HDPCG apparatus. *Protein Pept Lett.* 2015;22(2):173-9.
- Adachi S, Yoshizaki I, Ishikawa T, Yokoyama E, Furukawa Y, Shimaoka T. Stable growth mechanisms of ice disk crystals in heavy water. *Phys Rev E Stat Nonlin Soft Matter Phys.* 2011 Nov;84(5 Pt 1):051605. Epub 2011 Nov 22.
- Akdim MR, Goedheer WJ. Modeling of voids in colloidal plasmas. *Phys Rev E Stat Nonlin Soft Matter Phys.* 2002 Jan;65(1 Pt 2):015401. Epub 2001 Dec 11.
- Akdim MR, Goedheer WJ. Modeling the effect of dust on the plasma parameters in a dusty argon discharge under microgravity. *Phys Rev E Stat Nonlin Soft Matter Phys.* 2003 Jun;67(6 Pt 2):066407. Epub 2003 Jun 27.
- 5. Alvarado UR, DeWitt CR, Shultz BB, Ramsland PA, Edmundson AB. Crystallization of a human Bence–Jones protein in microgravity using vapor diffusion in capillaries. *Journal of Crystal Growth.* 2001;223(3):407-414.
- Aritake K, Kado Y, Inoue T, Miyano M, Urade Y. Structural and Functional Characterization of HQL-79, an Orally Selective Inhibitor of Human Hematopoietic Prostaglandin D Synthase. *J. Biol. Chem.* 2006 Jun 2;281(22):15277-15286. Epub 2006 Mar 17.
- 7. Asano K, Fujita S, Senda T, Mitsui Y. Crystal growth of ribonuclease S under microgravity. *J Cryst Growth*. 1992;122:323-9.

- Banco MT, Mishra V, Ostermann A, Schrader TE, Evans GB, Kovalevsky A, Ronning DR. Neutron structures of the Helicobacter pylori 5'-methylthioadenosine nucleosidase highlight proton sharing and protonation states. *Proc Natl Acad Sci U S A.* 2016 Nov 29; 113(48):13756-13761.
- Barnes CL, Snell EH, Kundrot CE. Thaumatin crystallization aboard the International Space Station using liquid-liquid diffusion in the Enhanced Gaseous Nitrogen Dewar (EGN). Acta Crystallogr D Biol Crystallogr. 2002 May;58(Pt 5):751-60. Epub 2002 Apr 26.
- Berisio R, Vitagliano L, Mazzarella L, Zagari A. Crystal structure of the collagen triple helix model [(Pro-Pro-Gly) (10)](3). *Protein Sci.* 2002 Feb;11(2):262-70.
- 11. Berisio R, Vitagliano L, Sorrentino G, Carotenuto L, Piccolo C, Mazzarella L, Zagari A. Effects of microgravity on the crystal quality of a collagen-like polypeptide. *Acta Crystallogr D Biol Crystallogr.* 2000 Jan;56(Pt 1):55-61.
- Berisio R, Vitagliano L, Vergara A, Sorrentino G, Mazzarella L, Zagari A. Crystallization of the collagen-like polypeptide (PPG)10 aboard the International Space Station. 2. Comparison of crystal quality by X-ray diffraction. *Acta Crystallogr D Biol Crystallogr.* 2002 Oct;58(Pt 10 Pt 1):1695-9. Epub 2002 Sep 26.
- Betzel C, Gunther N, Poll S, Moore K, DeLucas LJ, Bugg CE, Weber W. Crystallization of the EGF receptor ectodomain on US space mission STS-47. *Microgravity Sci Technol.* 1994 Sep;7(3):242-5.



- Betzel C, Gourinath S, Kumar P, Kaur P, Perbandt M, Eschenburg S, Singh TP. Structure of a serine protease proteinase K from Tritirachium album limber at 0.98 A resolution. *Biochemistry*. 2001 Mar 13;40(10):3080-3088.
- 15. Bi RC, Gui LL, Han Q, Shen FL, Shi K, Wang YP, Chen SZ, Hu YL, Niu XT, Dong J, Zhou YC, Lin NQ. Protein crystallization in space. *Microgravity Sci Technol.* 1994 Jul;7(2):203-6.
- Bi RC, Gui LL, Shi K, Wang YP, Chen SZ, Han Q, Hu YL, Shen FL, Niu XT, Hua ZQ, et al. Protein crystal growth in microgravity. Sci China B. 1994 Oct;37(10):1185-91.
- Blakeley MP, Langan P, Niimura N, Podjarny A. Neutron crystallography: opportunities, challenges, and limitations. *Current Opinion in Structural Biology.* 2008;18(5):593-600.
- Boggon TJ, Helliwell JR, Judge RA, Olczak A, Siddons DP, Snell EH, Stojanoff V. Synchrotron X-ray reciprocal-space mapping, topography and diffraction resolution studies of macromolecular crystal quality. *Acta Crystallogr D Biol Crystallogr.* 2000 Jul;56(Pt 7):868-80.
- Borgstahl GE, Vahedi-Faridi A, Lovelace J, Bellamy HD, Snell EH. A test of macromolecular crystallization in microgravity: large well ordered insulin crystals. *Acta Crystallogr D Biol Crystallogr*. 2001 Aug;57(Pt 8):1204-7. Epub 2001 Jul 23.
- Borisova SN, Birnbaum GI, Rose DR, Evans SV. Experiments in microgravity: a comparison of crystals of a carbohydrate-binding fab grown on the ground, on space shuttle Discovery and on space station Mir. Acta Crystallogr D Biol Crystallogr. 1996 Mar 1;52(Pt 2):267-71.
- 21. Bosch R, Lautenschlager P, Potthast L, Stapelmann J. Experiment equipment for protein crystallization in µg facilities. *Journal of Crystal Growth*. 1992;122(1-4):310-316.
- 22. Broutin-L'Hermite I, Ries-Kautt M, Ducruix A. 1.7 A X-ray structure of space-grown collagenase crystals. *Acta Crystallogr D Biol Crystallogr.* 2000 Mar 1;56(3):376-378.
- Evans CA, Robinson JA, Tate-Brown J, Thumm T, Crespo-Richey J, Baumann D, Rhatigan J. International Space Station Science Research Accomplishments During the Assembly Years: An Analysis of Results from 2000-2008. NASA/TP-2009-213146-Revision A. NASA Johnson Space Center, Houston, Tex.
- [Carter DC, Wright B, Miller T, Chapman J, Twigg P, Keeling K, Moody K, White M, James C, Ruble JR, Ho JX, Adcock-Downey L, Bunick G, Harp J. Diffusion-controlled crystallization apparatus for microgravity (DCAM): flight and ground-based applications. *Journal of Crystal Growth*. 1999;196(2-4):602-609.
- Carter DC, Wright B, Miller T, Chapman J, Twigg P, Keeling K, Moody K, White M, Click J, Ruble JR, Ho JX, Adcock-Downey L, Dowling T, Chang CH, Ala P, Rose J, Wang BC, Declercq JP, Evrard C, Rosenberg J, Wery JP, Clawson D, Wardell M, Stallings W, Stevens A. PCAM: a multi-user facility-based protein crystallization apparatus for microgravity. *Journal of Crystal Growth.* 1999;196(2-4):610-622.

- Castagnolo D, Piccolo C, Carotenuto L, Vergara A, Zagari A. Crystallization of the collagen-like polypeptide (PPG)10 aboard the International Space Station. 3. Analysis of residual acceleration-induced motion. *Acta Crystallogr D Biol Crystallogr.* 2003 Apr;59(Pt 4):773-6. Epub 2003 Mar 25.
- 27. Chayen NE, Gordon EJ, Zagalsky PF. Crystallization of apocrustacyanin on the International Microgravity Laboratory (IML-2) mission. *Acta Crystallogr D Biol Crystallogr.* 1996 Jan 1;52(Pt 1):156-9.
- 28. Chayen NE, Helliwell JR. Microgravity protein crystallization: are we reaping the full benefit of outer space? *Ann N Y Acad Sci.* 2002 Oct;974:591-7.
- Chayen, NE. Microgravity protein crystallisation aboard the photon satellite. *Journal of Crystal Growth*. 1995;153(3-4):175-179.
- Chen R, Liu J, Lu Q, Liu Y, Yin D. Effects of physical environments on nucleation of protein crystals: a review. *Sheng Wu Gong Cheng Xue Bao.* 2011 Jan;27(1):9-17. Review. Chinese.
- Cheng Z, Chaikin PM, Zhu J, Russel WB, Meyer WV. Crystallization kinetics of hard spheres in microgravity in the coexistence regime: interactions between growing crystallites. *Phys Rev Lett.* 2002 Jan 7;88(1):015501. Epub 2001 Dec 14.
- Cole T, Kathman A, Koszelak S, McPherson A. Determination of local refractive index for protein and virus crystals in solution by Mach-Zehnder interferometry. *Anal Biochem.* 1995 Oct 10;231(1):92-8.
- Kundrot CE, Judge RA, Pusey ML, Snell EH. Microgravity and Macromolecular Crystallography. *Cryst. Growth Des*. 2001;1(1):87-99.
- Dajnowicz S, Parks JM, Hu X, Gesler K, Kovalevsky AY, Mueser TC. Direct evidence that an extended hydrogenbonding network influences activation of pyridoxal 5 -phosphate in aspartate aminotransferase. *J Biol Chem.* 2017 Apr 7;292(14):5970-5980. doi: 10.1074/jbc. M116.774588. Epub 2017 Feb 23.
- 35. Day J, McPherson A. Macromolecular crystal growth experiments on International Microgravity Laboratory—1. *Protein Sci.* 1992 Oct;1(10):1254-68.
- Declercq JP, Evrard C, Lamzin V, Parello J. Crystal structure of the EF-hand parvalbumin at atomic resolution (0.91 A) and at low temperature (100 K). Evidence for conformational multistates within the hydrophobic core. *Protein Sci.* 1999 Oct;8(10):2194-204.
- Deisenhofer J, Epp O, Miki K, Huber R, Michel H. Structure of the protein subunits in the photosynthetic reaction centre of Rhodopseudomonas viridis at 3 Å resolution. *Nature*. 1985 Dec 19-1986 Jan1;318(6047):618-624.
- DeLucas LJ, Moore KM, Long MM. Protein crystal growth and the International Space Station. *Gravit Space Biol Bull.* 1999 May;12(2):39-45.



- DeLucas LJ, Smith CD, Carter DC, Twigg P, He XM, Snyder RS, Weber PC, Schloss JV, Einspahr HM, Clancy LL, McPherson A, Koszelak S, Vandonselaar MM, Prasad L, Quail JW, Delbaere LT, Bugg CE. Protein crystal growth aboard the U.S. space shuttle flights STS-31 and STS-32. *Adv Space Res.* 1992;12(1):393-400.
- DeLucas LJ, Smith CD, Smith HW, Vijay-Kumar S, Senadhi SE, Ealick SE, Carter DC, Snyder RS, Weber PC, Salemme FR, et al. Protein crystal growth in microgravity. Science. 1989 Nov 3;246(4930):651-4. *Erratum in: Science.* 1990 Sep 21;249(4975):1359.
- DeLucas LJ, Suddath FL, Snyder R, Naumann R, Broom MB, Pusey M, Yost V, Herren B, Carter D, Nelson B, Meehan EJ, McPherson A, Bugg CE. Preliminary investigations of protein crystal growth using the space shuttle. *Journal of Crystal Growth.* 1986;76(3):681-693.
- DeLucas LJ, Long MM, Moore KM, Rosenblum WM, Bray TL, Smith C, Carson M, Narayana SVL, Harrington MD, Carter D, Clark Jr AD, Nanni RG, Ding J, Jacobo-Molina A, Kamer G, Hughes SH, Arnold E, Einspahr HM, Clancy LL, Rao GSJ, Cook PF, Harris BG, Munson SH, Finzel BC, McPherson A, Weber PC, Lewandowski FA, Nagabhushan TL, Trotta PP, Reichert P, Navia MA, Wilson KP, Thomson JA, Richards RN, Bowersox KD, Meade CJ, Baker ES, Bishop SP, Dunbar BJ, Trinh E, Prahl J, Sacco Jr A, Bugg CE. Recent results and new hardware developments for protein crystal growth in microgravity. *Journal of Crystal Growth.* 1994;135(1-2):183-195.
- 43. Derby JJ, Kwon YI, Pandy A, Sonda P, Yeckel A, Jung T, Müller G. Developing quantitative, multiscale models for microgravity crystal growth. *Ann N Y Acad Sci.* 2006 Sep;1077:124-45.
- 44. Dobrianov I, Finkelstein KD, Lemay SG, Thorne RE.Xray topographic studies of protein crystal perfection and growth. *Acta Crystallogr D Biol Crystallogr.* 1998 Sep 1;54(Pt 5):922-37.
- Dong J, Boggon TJ, Chayen NE, Raftery J, Bi RC, Helliwell JR. Bound-solvent structures for microgravity-, ground control-, gel- and microbatch-grown hen egg-white lysozyme crystals at 1.8 A resolution. *Acta Crystallogr D Biol Crystallogr.* 1999 Apr;55(Pt 4):745-52.
- 46. Duan L, Kang Q, Hu WR, Li GP, Wang DC. The mass transfer process and the growth rate of protein crystals. *Biophys Chem.* 2002 Jun 19;97(2-3):189-201.
- Ealick SE, Babu YS, Bugg CE, Erion MD, Guida WC, Montgomery JA, Secrist JA. Application of crystallographic and modeling methods in the design of purine nucleoside phosphorylase inhibitors. *Proceedings of the National Academy of Sciences of the United States of America*. 1991;88(24):11540-11544.
- Ealick S, Cook W, Vijay-Kumar S, Carson M, Nagabhushan T, Trotta P, Bugg C. Three-dimensional structure of recombinant human interferon- gamma. *Science*. 1991;252(5006):698-702.
- 49. Esposito L, Sica F, Sorrentino G, Berisio R, Carotenuto L, Giordano A, Raia CA, Rossi M, Lamzin VS, Wilson KS,

Zagari A. Protein crystal growth in the Advanced Protein Crystallization Facility on the LMS mission: a comparison of Sulfolobus solfataricus alcohol dehydrogenase crystals grown on the ground and in microgravity. *Acta Crystallogr D Biol Crystallogr.* 1998 May 1;54(Pt 3):386-90.

- Esposito L, Sica F, Raia CA, Giordano A, Rossi M, Mazzarella L, Zagari A. Crystal Structure of the Alcohol Dehydrogenase from the Hyperthermophilic Archaeon Sulfolobus solfataricus at 1.85 Å Resolution. *Journal of Molecular Biology.* 2002;318(2):463-477.
- 51. Evrard C, Fastrez J, Declercq JP. Crystal structure of the lysozyme from bacteriophage lambda and its relationship with V and C-type lysozymes1. *Journal of Molecular Biology.* 1998 Feb 13;276(1):151-164.
- 52. Fromme P, Grotjohann I. Crystallization of Photosynthetic Membrane Proteins. *Current Topics in Membranes*. 2009;63:191-227.
- 53. Furukawa Y, Nagashima K, Nakatsubo SI, Yoshizaki I, Tamaru H, Shimaoka T, Sone T, Yokoyama E, Zepeda S, Terasawa T, Asakawa H, Murata KI, Sazaki G. Oscillations and accelerations of ice crystal growth rates in microgravity in presence of antifreeze glycoprotein impurity in supercooled water. *Sci Rep.* 2017 Mar 6;7:43157. doi: 10.1038/srep43157.
- 54. Galkin O, Vekilov PG. Control of protein crystal nucleation around the metastable liquid-liquid phase boundary. *Proc Natl Acad Sci U S A.* 2000 Jun 6;97(12): 6277-81.
- Gerdts CJ, Elliott M, Lovell S, Mixon MB, Napuli AJ, Staker BL, Nollert P, Stewart L. The plug-based nanovolume Microcapillary Protein Crystallization System (MPCS). Acta Crystallogr D Biol Crystallogr. 2008;64(Pt 11):1116-1122.
- Gilliland GL, Tung M, Ladner J. The Biological Macromolecule Crystallization Database and NASA Protein Crystal Growth Archive. *J Res Natl Inst Stand Technol.* 1996 May-Jun;101(3):309-20.
- Gonzalez F, Cunisse M, Perigaud A. CROCODILE: an automated apparatus for organic crystal growth from solution. *Acta Astronaut.* 1991 Dec;25(12):775-84.
- Gonzalez-Ramirez LA, Carrera J, Gavira JA, Melero-Garcia E, Garcia-Ruiz JM. Granada Crystallization Facility-2: A Versatile Platform for Crystallization in Space. *Crystal Growth & Design.* 2008;8(12):4324-4329.
- 59. Goree J, Morfill GE, Tsytovich VN, Vladimirov SV. Theory of dust voids in plasmas. *Phys Rev E Stat Phys Plasmas Fluids Relat Interdiscip Topics*. 1999 Jun;59(6):7055-67.
- Gourinath S, Degenhardt M, Eschenburg S, Moore K, Delucas LJ, Betzel C, Singh TP. Mercury induced modifications in the stereochemistry of the active site through Cys-73 in a serine protease--crystal structure of the complex of a partially modified proteinase K with mercury at 1.8 A resolution. *Indian J Biochem Biophys.* 2001 Oct;38(5):298-302.
- 61. Grabowski, M., M. Chruszcz, M. D. Zimmerman, O. Kirillova and W. Minor. Benefits of Structural Genomics for Drug Discovery Research. *Infectious Disorders Drug Targets.* 2009;9(5): 459-474.



- 62. Gupta NR, Haj-Hariri H, Borhan A. Double-layer thermocapillary convection in a differentially heated cavity. *Ann N Y Acad Sci.* 2006 Sep;1077:395-414.
- 63. Habash J, Boggon TJ, Raftery J, Chayen NE, Zagalsky PF, Helliwell JR. Apocrustacyanin C(1) crystals grown in space and on earth using vapour-diffusion geometry: protein structure refinements and electron-density map comparisons. *Acta Crystallogr D Biol Crystallogr.* 2003 Jul;59(Pt 7):1117-23. Epub 2003 Jun 27. Review.
- 64. Harp JM, Hanson BL, Timm DE, Bunick GJ. Asymmetries in the nucleosome core particle at 2.5 A resolution. *Acta Crystallographica Section D.* 2000 Dec;56(12):1513-1534.
- 65. He XM, Carter DC. Atomic structure and chemistry of human serum albumin. *Nature.* 1992 Jul 16;358(6383):209-215.
- 66. Hirschler J, Fontecilla-Camps JC. Contaminant effects on protein crystal morphology in different growth environments. *Acta Crystallogr D Biol Crystallogr.* 1996 Jul 1;52(Pt 4):806-12.
- 67. Ho JX, Declercq JP, Myles DAA, Wright BS, Ruble JR, Carter DC. Neutron structure of monoclinic lysozyme crystals produced in microgravity. *Journal of Crystal Growth.* 2001;232(1-4):317-325.
- Inaka K, Takahashi S, Aritake K, Tsurumura T, Furubayashi N, Yan B, Hirota E, Sano S, Sato M, Kobayashi T, Yoshimura Y, Tanaka H, Urade Y. High-Quality Protein Crystal Growth of Mouse Lipocalin-Type Prostaglandin D Synthase in Microgravity. *Cryst Growth Des.* 2011 Jun 1;11(6):2107-2111. Epub 2011 Apr 5.
- Judge RA, Forsythe EL, Pusey ML. The effect of protein impurities on lysozyme crystal growth. *Biotechnol Bioeng.* 1998 Sep 20;59(6):776-85.
- Judge RA, Jacobs RS, Frazier T, Snell EH, Pusey ML. The effect of temperature and solution pH on the nucleation of tetragonal lysozyme crystals. *Biophys J.* 1999 Sep;77(3):1585-93.
- Jun D, Yaoping W, Qing H, Ruchang B. Structural study on hen egg-white lysozyme crystals grown in gravity and microgravity. *Sci China C Life Sci.* 1998 Jun;41(3):238-44. doi: 10.1007/BF02895097.
- 72. Kahsay RY, Gao G, Liao L. An improved hidden Markov model for transmembrane protein detection and topology prediction and its applications to complete genomes. *Bioinformatics.* 2005;21(9):1853-1858.
- Khurshid S, Chayen NE. Upside-down protein crystallization: designing microbatch experiments for microgravity. *Ann N Y Acad Sci.* 2006 Sep;1077:208-13.
- Kinoshita T, Hashimoto T, Sogabe Y, Fukada H, Matsumoto T, Sawa M. High-resolution structure discloses the potential for allosteric regulation of mitogen-activated protein kinase kinase 7. *Biochem Biophys Res Commun.* 2017 Sep 7. pii: S0006-291X(17)31785-0. doi: 10.1016/j.bbrc.2017.09.025. [Epub ahead of print]

- 75. Kinoshita T, Maruki R, Warizaya M, Nakajima H, Nishimura S. Structure of a high-resolution crystal form of human triosephosphate isomerase: improvement of crystals using the gel-tube method. *Acta Crystallogr Sect F Struct Biol Cryst Commun.* 2005 Apr 1;61(Pt 4):346-9. Epub 2005 Mar 24.
- Klukas O, Schubert WD, Jordan P, Krauß N, Fromme P, Witt HT, Saenger W. Photosystem I, an Improved Model of the Stromal Subunits PsaC, PsaD, and PsaE. *J. Biol. Chem.* 1999; 274(11):7351-7360.
- Ko TP, Day J, McPherson A. The refined structure of canavalin from jack bean in two crystal forms at 2.1 and 2.0 A resolution. *Acta Crystallogr D Biol Crystallogr.* 2000;56(Pt 4):411-420.
- Ko TP, Day J, Malkin AJ, McPherson A. Structure of orthorhombic crystals of beef liver catalase. *Acta Crystallographica Section D.* 1999;55(8):1383-1394.
- 79. Koszelak S, Day J, Leja C, Cudney R, McPherson A. Protein and virus crystal growth on international microgravity laboratory-2. *Biophys J.* 1995 Jul;69(1):13-9.
- Koszelak S, Leja C, McPherson A. Crystallization of Biological Macromolecules from Flash Frozen Samples on the Russian Space Station Mir. *Biotechnology and Bioengineering*. 1996;2:449-458.
- Krauspenhaar R, Rypniewski W, Kalkura N, Moore K, DeLucas L, Stoeva S, Mikhailov A, Voelter W, Betzel Ch. Crystallisation under microgravity of mistletoe lectin I from Viscum album with adenine monophosphate and the crystal structure at 1.9 A resolution. *Acta Crystallogr D Biol Crystallogr.* 2002 Oct;58(Pt 10 Pt 1):1704-7. Epub 2002 Sep 26.
- Lappa M, Carotenuto L. Effect of convective disturbances induced by g-jitter on the periodic precipitation of lysozyme. *Microgravity Sci Technol.* 2003;14(2):41-56.
- Lappa M, Piccolo C, Carotenuto L. Numerical and experimental analysis of periodic patterns and sedimentation of lysozyme. *J Cryst Growth.* 2003 Jul;254(3-4):469-86.
- Lappa M. The growth and the fluid dynamics of protein crystals and soft organic tissues: models and simulations, similarities and differences. *J Theor Biol.* 2003 Sep 21;224(2):225-40.
- Larson SB, Day JS, McPherson A. Satellite tobacco mosaic virus refined to 1.4 Å resolution. *Acta Crystallogr D Biol Crystallogr.* 2014 Sep;70(Pt 9):2316-30. doi: 10.1107/ S1399004714013789. Epub 2014 Aug 29.
- Larson SB, Day J, Greenwood A, McPherson A. Refined structure of satellite tobacco mosaic virus at 1.8 Å resolution. *Journal of Molecular Biology.* 1998 Mar 20;277(1):37-59.
- Li S, Yang S, Du Q, Chen H, Lin Z. The pre-experiment on space crystallization of pig heart mitochondrial F1-ATPase. Space Med Med Eng (Beijing). 1998 Oct;11(5):343-6. Chinese.



- Lin H, Petsev DN, Yau ST, Thomas BR, Vekilov PG. Lower Incorporation of Impurities in Ferritin Crystals by Suppression of Convection: Modeling Results. *Crystal Growth & Design.* 2001;1(1):73-79.
- Lin H, Rosenberger F, Alexander JID, Nadarajah A. Convective-diffusive transport in protein crystal growth. *Journal of Crystal Growth*. 1995;151(1-2):153-162.
- 90. Littke W, John C. Materials: Protein single crystal growth under microgravity. *Journal of Crystal Growth*. 1986;76(3):663-672.
- Long MM, DeLucas LJ, Smith C, Carson M, Moore K, Harrington MD, Pillion DJ, Bishop SP, Rosenblum WM, Naumann RJ, Chait A, Prahl J, Bugg CE. Protein crystal growth in microgravity-temperature induced large scale crystallization of insulin. *Microgravity Sci Technol.* 1994 Jul;7(2):196-202.
- 92. Long MM, Bishop JB, Nagabhushan TL, Reichert P, Smith GD, DeLucas LJ. Protein crystal growth in microgravity review of large scale temperature induction method: bovine insulin, human insulin and human alpha interferon. *Journal of Crystal Growth.* 1996;168(1-4):233-243.
- Lorber B, Théobald-Dietrich A, Charron C, Sauter C, Ng JD, Zhu DW, Giegé R. From conventional crystallization to better crystals from space: a review on pilot crystallogenesis studies with aspartyl-tRNA synthetases. *Acta Crystallogr D Biol Crystallogr.* 2002 Oct;58(Pt 10 Pt 1):1674-80. Epub 2002 Sep 26. Review.
- Lorber, B. The crystallization of biological macromolecules under microgravity: a way to more accurate threedimensional structures? *Biochimica et Biophysica Acta* (*BBA*) - *Proteins and Proteomics*. 2002;1599(1-2):1-8.
- Lu HM, Yin DC, Li HS, Geng LQ, Zhang CY, Lu QQ, Guo YZ, Guo WH, Shang P, Wakayama NI. A containerless levitation setup for liquid processing in a superconducting magnet. *Rev Sci Instrum.* 2008 Sep;79(9):093903. doi: 10.1063/1.2980383.
- Małecki PH, Rypniewski W, Szymański M, Barciszewski J, Meyer A. Binding of the plant hormone kinetin in the active site of Mistletoe Lectin I from Viscum album. *Biochim Biophys Acta*. 2012 Feb;1824(2):334-8. doi: 10.1016/j.bbapap.2011.10.013. Epub 2011 Oct 28.
- Maes D, Evrard C, Gavira JA, Sleutel M, Van De Weerdt C, Otalora F, Garcia-Ruiz JM, Nicolis G, Martial J, Decanniere K. Toward a Definition of X-ray Crystal Quality. *Crystal Growth & Design.* 2008;8(12):4284-4290.
- 98. McPherson A, DeLucas L. Crystal-growing in space. *Science*. 1999 Mar 5;283(5407):1459.
- 99. McPherson A, DeLucas LJ. Microgravity Protein Crystallization. *NPJ Microgravity*. 2015 Sep 3;1:15010.
- McPherson A, Malkin AJ, Kuznetsov YG, Koszelak S, Wells M, Jenkins G, Howard J, Lawson G. The effects of microgravity on protein crystallization: evidence for concentration gradients around growing crystals. *Journal of Crystal Growth.* 1999;196(2-4):572-586.

- Meyer A, Rypniewski W, Szymański M, Voelter W, Barciszewski J, Betzel C. Structure of mistletoe lectin I from Viscum album in complex with the phytohormone zeatin. *Biochim Biophys Acta*. 2008 Nov;1784(11):1590-5. doi: 10.1016/j.bbapap.2008.07.010. Epub 2008 Jul 31.
- 102. Miele AE, Federici L, Sciara G, Draghi F, Brunori M, Vallone B. Analysis of the effect of microgravity on protein crystal quality: the case of a myoglobin triple mutant. *Acta Crystallogr D Biol Crystallogr.* 2003 Jun;59(Pt 6):982-8. Epub 2003 May 23.
- Mohamad Aris SN, Thean Chor AL, Mohamad Ali MS, Basri M, Salleh AB, Raja Abd Rahman RN. Crystallographic analysis of ground and space thermostable T1 lipase crystal obtained via counter diffusion method approach. *Biomed Res Int.* 2014;2014:904381. doi: 10.1155/2014/904381. Epub 2014 Jan 2.
- Nanev CN, Penkova A, Chayen N. Effects of buoyancydriven convection on nucleation and growth of protein crystals. Ann N Y Acad Sci. 2004 Nov;1027:1-9.
- Narayana SVL, Carson M, el-Kabbani O, Kilpatrick JM, Moore D, Chen X, Bugg CE, Volanakis JE, DeLucas LJ. Structure of Human Factor D: Complement System Protein at 2.0 Å Resolution. J. Mol. Biol. 1994;235(2):695-708.
- 106. Ng JD, Baird JK, Coates L, Garcia-Ruiz JM, Hodge TA, Huang S. Large-volume protein crystal growth for neutron macromolecular crystallography. Acta Crystallogr F Struct Biol Commun. 2015 Apr;71(Pt 4):358-70. doi: 10.1107/ S2053230X15005348. Epub 2015 Mar 30.
- Ng JD, Lorber B, Giege R, Koszelak S, Day J, Greenwood A, McPherson A. Comparative analysis of thaumatin crystals grown on earth and in microgravity. *Acta Crystallogr D Biol Crystallogr.* 1997 Nov 1;53(Pt 6):724-33.
- 108. Ng JD, Sauter C, Lorber B, Kirkland N, Arnez J, Giegé R. Comparative analysis of space-grown and earth-grown crystals of an aminoacyl-tRNA synthetase: space-grown crystals are more useful for structural determination. *Acta Crystallogr D Biol Crystallogr.* 2002 Apr;58(Pt 4):645-52. Epub 2002 Mar 22.
- 109. Ng JD, Baird JK, Coates L, Garcia-Ruiz JM, Hodge TA, Huang S. Large-volume protein crystal growth for neutron macromolecular crystallography. *Acta Crystallogr F Struct Biol Commun.* 2015 Apr;71(Pt 4):358-370.
- 110. Niimura N, Kurihara K, Ataka M. Dissolution rate of hen egg-white lysozyme crystal under microgravity. *Biol Sci Space.* 2001 Oct;15 Suppl:S176.
- 111. Niimura N, Bau R. Neutron protein crystallography: beyond the folding structure of biological macromolecules. *Acta Crystallogr A.* 2008;64(Pt 1):12-22.
- 112. Norvell JC, Berg JM. Update on the Protein Structure Initiative. *Structure*. 2007 Dec;15(12):1519-1522.
- 113. Ockels WJ. The first parabolic flight campaign for students. *ESA Bull.* 1995 May;82:4 p..



- 114. Oda K, Matoba Y, Noda M, Kumagai T, Sugiyama M. Catalytic mechanism of bleomycin N-acetyltransferase proposed on the basis of its crystal structure. *J Biol Chem.* 2010 Jan 8;285(2):1446-56. doi: 10.1074/jbc. M109.022277. Epub 2009 Nov 3.
- 115. Okutani T, Nagai H, Mamiya M, Shibuya M, Castillo M. Effect of microgravity and magnetic field on the metallic and crystalline structure of magnetostrictive SmFe2 synthesized by unidirectional solidification. *Ann N Y Acad Sci.* 2006 Sep;1077:146-60.
- 116. Okutani T, Nagai H, Mamiya M. Synthesis of highperformance magnetostrictive Tb0.3Dy0.7Fe2 by unidirectional solidification in microgravity. *Ann N Y Acad Sci.* 2009 Apr;1161:437-51. doi: 10.1111/j.1749-6632.2009.04084.x.
- Otálora F, Capelle B, Ducruix A, García-Ruiz JM. Mosaic spread characterization of microgravity-grown tetragonal lysozyme single crystals. *Acta Crystallogr D Biol Crystallogr.* 1999 Mar;55(Pt 3):644-9.
- Otálora F, García-Ruiz JM, Carotenuto L, Castagnolo D, Novella ML, Chernov AA. Lysozyme crystal growth kinetics in microgravity. *Acta Crystallogr D Biol Crystallogr.* 2002 Oct;58(Pt 10 Pt 1):1681-9. Epub 2002 Sep 26.
- 119. Otálora F, Gavira JA, Ng JD, García-Ruiz JM. Counterdiffusion methods applied to protein crystallization. *Prog Biophys Mol Biol.* 2009 Nov;101(1-3):26-37. doi: 10.1016/j.pbiomolbio.2009.12.004. Epub 2009 Dec 16. Review.
- 120. Otálora F, Novella ML, Gavira JA, Thomas BR, García-Ruiz JM. Experimental evidence for the stability of the depletion zone around a growing protein crystal under microgravity. *Acta Crystallogr D Biol Crystallogr.* 2001 Mar;57(Pt 3):412-7.
- 121. Overington JP, Al-Lazikani B, Hopkins AL. How many drug targets are there? *Nat Rev Drug Discov.* 2006;5(12):993-996.
- 122. Owen RB, Kroes RL, Witherow WK. Results and further experiments using Spacelab holography. *Opt Lett.* 1986 Jul 1;11(7):407-9.
- 123. Owen RB, Zozulya AA. Comparative study with doubleexposure digital holographic interferometry and a shackhartmann sensor to characterize transparent materials. *Appl Opt.* 2002 Oct 1;41(28):5891-5.
- 124. Owens GE, New DM, Olvera AI, Manzella JA, Macon BL, Dunn JC, Cooper DA, Rouleau RL, Connor DS, Bjorkman PJ .Comparative analysis of anti-polyglutamine Fab crystals grown on Earth and in microgravity. Acta Crystallogr F Struct Biol Commun. 2016 Oct 1;72(Pt 10):762-771. Epub 2016 Sep 22.
- 125. Owens GE, New DM, West AP Jr, Bjorkman PJ. Anti-PolyQ Antibodies Recognize a Short PolyQ Stretch in Both Normal and Mutant Huntingtin Exon 1. *J Mol Biol.* 2015 Jul 31; 427(15):2507-19. doi: 10.1016/j. jmb.2015.05.023. Epub 2015 Jun 3.
- 126. Patiño-Lopez L, Decanniere K, Gavira J, Maes D, Otalora F. Protein Experiment: Scientific Data Processing Platform for On-Flight Experiment Tuning. *Microgravity Science and Technology.* 2012 Nov;24(5):327-334.

- 127. Penkova A, Pan W, Hodjaoglu F, Vekilov PG. Nucleation of protein crystals under the influence of solution shear flow. *Ann N Y Acad Sci.* 2006 Sep;1077:214-31.
- 128. Pereda J, Mota FL, Chen L, Billia B, Tourret D, Song Y, Debierre JM, Guérin R, Karma A, Trivedi R, Bergeon N. Experimental observation of oscillatory cellular patterns in three-dimensional directional solidification. *Phys Rev E*. 2017 Jan;95(1-1):012803. doi: 10.1103/ PhysRevE.95.012803. Epub 2017 Jan 13.
- 129. Phan SE, Li M, Russel WB, Zhu J, Chaikin PM, Lant CT. Linear viscoelasticity of hard sphere colloidal crystals from resonance detected with dynamic light scattering. *Phys Rev E Stat Phys Plasmas Fluids Relat Interdiscip Topics.* 1999 Aug;60(2 Pt B):1988-98.
- 130. Pletser V, Bosch R, Potthast L, Lautenschlager P, Kassel R. The Protein Crystallisation Diagnostics Facility (PCDF) on Board ESA Columbus Laboratory. *Microgravity Science and Technology*. 2009;21(3):269-277.
- Prasad L, Sharma S, Vandonselaar M, Quail JW, Lee JS, Waygood EB, Wilson KS, Dauter Z, Delbaere LT. Evaluation of mutagenesis for epitope mapping. Structure of an antibody-protein antigen complex. *J Biol Chem.* 1993;268(15):10705-10708.
- 132. Pusey M, Witherow W, Naumann R. Preliminary investigations into solutal flow about growing tetragonal lysozyme crystals. *Journal of Crystal Growth.* 1988;90(1-3):105-111.
- Sabin J, Bailey AE, Espinosa G, Frisken BJ. Crystalarrested phase separation. *Phys Rev Lett.* 2012 Nov 9;109(19):195701. Epub 2012 Nov 7.
- 134. Safonova TN, Mikhailov SN, Veiko VP, Mordkovich NN, Manuvera VA, Alekseev CS, Kovalchuk MV, Popov VO, Polyakov KM. High-syn conformation of uridine and asymmetry of the hexameric molecule revealed in the high-resolution structures of Shewanella oneidensis MR-1 uridine phosphorylase in the free form and in complex with uridine. *Acta Crystallogr D Biol Crystallogr.* 2014 Dec 1;70(Pt 12):3310-9. doi: 10.1107/ S1399004714024079. Epub 2014 Nov 28.
- 135. Safonova TN, Mordkovich NN, Polyakov KM, Manuvera VA, Veiko VP, Popov VO. Crystallization of uridine phosphorylase from Shewanella oneidensis MR-1 in the laboratory and under microgravity and preliminary X-ray diffraction analysis. *Acta Crystallogr Sect F Struct Biol Cryst Commun.* 2012 Nov 1;68(Pt 11):1387-9. doi: 10.1107/ S1744309112041784. Epub 2012 Oct 30.
- 136. Sauter C, Otalora F, Gavira JA, Vidal O, Giege R, Garcia-Ruiz JM. Structure of tetragonal hen egg-white lysozyme at 0.94 A from crystals grown by the counter-diffusion method. *Acta Crystallogr D Biol Crystallogr.* 2001;57(Pt 8):1119-1126.
- 137. Sjölin L, Wlodawer A, Bergqvist G, Holm P, Loth K, Malmström H, Zaar J, Svensson LA, Gilliland GL. Protein crystal growth of Ribonuclease A and Pancreatic Trypsin Inhibitor aboard the MASER 3 rocket. *Journal of Crystal Growth.* 1991;110(1-2):322-332



- 138. Skinner R, Abrahams JP, Whisstock JC, Lesk AM, Carrell RW, Wardell MR. The 2.6 A structure of antithrombin indicates a conformational change at the heparin binding site. *J Mol Biol.* 1997 Feb 28;266(3):601-9.
- 139. Smith GD, Ciszak E, Pangborn W. A novel complex of a phenolic derivative with insulin: Structural features related to the T R transition. *Protein Science*. 1996;5:1502-1511.
- 140. Smith GD, Pangborn WA, Blessing RH. The structure of T6 human insulin at 1.0 A resolution. *Acta Crystallogr D Biol Crystallogr.* 2003;59(Pt 3):474-482.
- 141. Snell EH, Boggon TJ, Helliwell JR, Moskowitz ME, Nadarajah A. CCD video observation of microgravity crystallization of lysozyme and correlation with accelerometer data. *Acta Crystallogr D Biol Crystallogr.* 1997 Nov 1;53(Pt 6):747-55.
- 142. Snell EH, Cassetta A, Helliwell JR, Boggon TJ, Chayen NE, Weckert E, Hölzer K, Schroer K, Gordon EJ, Zagalsky PF. Partial improvement of crystal quality for microgravitygrown apocrustacyanin C1. Acta Crystallogr D Biol Crystallogr. 1997 May 1;53(Pt 3):231-9.
- 143. Snell EH, Helliwell JR, Boggon TJ, Lautenschlager P, Potthast L. Lysozyme crystal growth kinetics monitored using a Mach-Zehnder interferometer. *Acta Crystallogr D Biol Crystallogr.* 1996 May 1;52(Pt 3):529-33.
- 144. Snell EH, Weisgerber S, Helliwell JR, Hölzer K, Schroer K. Improvements in lysozyme protein crystal perfection through microgravity growth. *Acta Crystallogr D Biol Crystallogr.* 1995 Nov 1;51(Pt 6):1099-102.
- Snell EH, Helliwell JR. Macromolecular crystallization in microgravity. *Reports on Progress in Physics*. 2005;68:799-853.
- 146. Snyder RS, Fuhrmann K, Walter HU. Protein crystallization facilities for microgravity experiments. *Journal of Crystal Growth.* 1991; 110(1-2):333-338.
- 147. Someya S, Munakata T, Nishio M, Okamoto K, Madarame H. Flow characteristics of two immiscible liquid layers subjected to a horizontal temperature gradient. *Ann N Y Acad Sci.* 2002 Oct;972:299-306.
- 148. Steiner B, Dobbyn RC, Black D, Burdette H, Kuriyama M, Spal R, van den Berg L, Fripp A, Simchick R, Lal RB, Batra A, Matthiesen D, Ditchek B. High Resolution Synchrotron X-Radiation Diffraction Imaging of Crystals Grown in Microgravity and Closely Related Terrestrial Crystals. J Res Natl Inst Stand Technol. 1991 May-Jun;96(3):305-331. doi: 10.6028/jres.096.017.
- 149. Stoddard BL, Strong RK, Farber GK, Arrott A, Petsko GA. Design of apparatus and experiments to determine the effect of microgravity on the crystallization of biological macromolecules using the MIR spacestation. *Journal of Crystal Growth*. 1991;110(1-2):312-316.
- 150. Sun B, Sun Z, Ouyang W, Xu S. Structural ordering and glass forming of soft spherical particles with harmonic repulsions. *J Chem Phys.* 2014 Apr 7;140(13):134904. doi: 10.1063/1.4869833.

- 151. Suvorova EI, Christensson F, Lundager Madsen HE, Chernov AA. Terrestrial and space-grown HAP and OCP crystals: effect of growth conditions on perfection and morphology. *J Cryst Growth.* 1998 Mar 1;186(1-2):262-74.
- Sygusch J, Coulombe R, Cassanto JM, Sportiello MG, Todd P. Protein crystallization in low gravity by step gradient diffusion method. *J Cryst Growth.* 1996 May 1;162(3-4):167-72.
- Symersky J, Devedjiev Y, Moore K, Brouillette C, DeLucas L. NH3-dependent NAD+ synthetase from Bacillus subtilis at 1 A resolution. *Acta Crystallogr D Biol Crystallogr.* 2002 Jul;58(Pt 7):1138-46. Epub 2002 Jun 20.
- 154. Takahashi S, Ohta K, Furubayashi N, Yan B, Koga M, Wada Y, Yamada M, Inaka K, Tanaka H, Miyoshi H, Kobayashi T, Kamigaichi S. JAXA protein crystallization in space: ongoing improvements for growing high-quality crystals. *J Synchrotron Radiat*. 2013 Nov;20(Pt 6):968-73. doi: 10.1107/S0909049513021596. Epub 2013 Sep 26.
- 155. Takahashi S, Tsurumura T, Aritake K, Furubayashi N, Sato M, Yamanaka M, Hirota E, Sano S, Kobayashi T, Tanaka T, Inaka K, Tanaka H, Urade Y. High-quality crystals of human haematopoietic prostaglandin D synthase with novel inhibitors. *Acta Crystallogr Sect F Struct Biol Cryst Commun.* 2010 Jul 1;66(Pt 7):846-50. doi: 10.1107/S1744309110020828. Epub 2010 Jun 24.
- 156. Tanaka H, Inaka K, Sugiyama S, Takahashi S, Sano S, Sato M, Yoshitomi S. Numerical analysis of the depletion zone formation around a growing protein crystal. *Ann N Y Acad Sci.* 2004 Nov;1027:10-9.
- 157. Tanaka H, Sasaki S, Takahashi S, Inaka K, Wada Y, Yamada M, Ohta K, Miyoshi H, Kobayashi T, Kamigaichi S. Numerical model of protein crystal growth in a diffusive field such as the microgravity environment. *J Synchrotron Radiat.* 2013 Nov;20(Pt 6):1003-9. doi: 10.1107/ S0909049513022784. Epub 2013 Oct 1.
- 158. Tanaka H, Umehara T, Inaka K, Takahashi S, Shibata R, Bessho Y, Sato M, Sugiyama S, Fusatomi E, Terada T, Shirouzu M, Sano S, Motohara M, Kobayashi T, Tanaka T, Tanaka A, Yokoyama S. Crystallization of the archaeal transcription termination factor NusA: a significant decrease in twinning under microgravity conditions. *Acta Crystallogr Sect F Struct Biol Cryst Commun.* 2007 Feb 1;63(Pt 2):69-73. Epub 2007 Jan 17.
- 159. Tanaka, H., T. Tsurumura, K. Aritake, N. Furubayashi, S. Takahashi, M. Yamanaka, E. Hirota, S. Sano, M. Sato, T. Kobayashi, T. Tanaka, K. Inaka and Y. Urade. Improvement in the quality of hematopoietic prostaglandin D synthase crystals in a microgravity environment. *Journal of Synchrotron Radiation*. 2011 Jan 1;18(1):88-91.
- 160. Terzyan SS, Bourne CR, Ramsland PA, Bourne PC, Edmundson AB. Comparison of the three-dimensional structures of a human Bence-Jones dimer crystallized on Earth and aboard US Space Shuttle Mission STS-95. *J Mol Recognit.* 2003 Mar-Apr;16(2):83-90.



- Thomas BR, Vekilov PG, Rosenberger F. Effects of microheterogeneity in hen egg-white lysozyme crystallization. *Acta Crystallogr D Biol Crystallogr.* 1998 Mar 1;54(Pt 2):226-36.
- 162. Thomas BR, Vekilov PG, Rosenberger F. Heterogeneity determination and purification of commercial hen eggwhite lysozyme. *Acta Crystallogr D Biol Crystallogr.* 1996 Jul 1;52(Pt 4):776-84.
- 163. Timofeev V, Slutskaya E, Gorbacheva M, Boyko K, Rakitina T, Korzhenevskiy D, Lipkin A, Popov V. Structure of recombinant prolidase from Thermococcus sibiricus in space group P21221. Acta Crystallogr F Struct Biol Commun. 2015 Aug;71(Pt 8):951-7. doi: 10.1107/ S2053230X15009498. Epub 2015 Jul 28.
- 164. Timofeev VI, Chuprov-Netochin RN, Samigina VR, Bezuglov VV, Miroshnikov KA, Kuranova IP. X-ray investigation of gene-engineered human insulin crystallized from a solution containing polysialic acid. Acta Crystallogr Sect F Struct Biol Cryst Commun. 2010 Mar 1;66(Pt 3):259-63. doi: 10.1107/S1744309110000461. Epub 2010 Feb 23.
- 165. Todd P, Vellinger JC, Sengupta S, Sportiello MG, Greenberg AR, Krantz WB. Sliding-cavity fluid contactors in lowgravity fluids, materials, and biotechnology research. *Ann N Y Acad Sci*. 2002 Oct;974:581-90.
- 166. Trakhanov SD, Grebenko AI, Shirokov VA, Gudkov AV, Egorov AV, Barmin IN, Vainstein BK, Spirin AS. Crystallization of protein and ribosomal particles in microgravity. *Journal of Crystal Growth*. 1991;110(1-2):317-321.
- Trolinger JD, Lal RB, McIntosh D, Witherow WK. Holographic particle-image velocimetry in the first International Microgravity Laboratory aboard the Space Shuttle Discovery. *Appl Opt.* 1996 Feb 1;35(4):681-9. doi: 10.1364/A0.35.000681.
- Tsige M, Mahajan MP, Rosenblatt C, Taylor PL. Nematic order in nanoscopic liquid crystal droplets. *Phys Rev E Stat Phys Plasmas Fluids Relat Interdiscip Topics*. 1999 Jul;60(1):638-44.
- Tuszynski JA, Sataric MV, Portet S, Dixon JM. Gravitational symmetry breaking leads to a polar liquid crystal phase of microtubules in vitro. *J Biol Phys.* 2005 Dec;31(3-4):477-86. doi: 10.1007/s10867-005-7284-5.
- 170. Vahedi-Faridi A, Lovelace J, Bellamy HD, Snell EH, Borgstahl GE. Physical and structural studies on the cryocooling of insulin crystals. *Acta Crystallogr D Biol Crystallogr.* 2003 Dec;59(Pt 12):2169-82. Epub 2003 Nov 27.
- 171. Vahedi-Faridi A, Porta J, Borgstahl GE. Improved threedimensional growth of manganese superoxide dismutase crystals on the International Space Station. *Acta Crystallogr D Biol Crystallogr.* 2003 Feb;59(Pt 2):385-8. Epub 2003 Jan 23.
- 172. Vallazza M, Banumathi S, Perbandt M, Moore K, DeLucas L, Betzel C, Erdmann VA. Crystallization and structure analysis of Thermus flavus 5S rRNA helix B. Acta Crystallogr D Biol Crystallogr. 2002 Oct;58(Pt 10 Pt 1):1700-3. Epub 2002 Sep 26.

- 173. van der Woerd MJ, Ferree D, Pusey ML. The promise of macromolecular crystallization in micrfluidic chips. *Journal of Structural Biology*. 2003;142(1):180-187.
- 174. Vaney MC, Maignan S, Riès-Kautt M, Ducriux A. High-resolution structure (1.33 A) of a HEW lysozyme tetragonal crystal grown in the APCF apparatus. Data and structural comparison with a crystal grown under microgravity from SpaceHab-01 mission. *Acta Crystallogr D Biol Crystallogr.* 1996 May 1;52(Pt 3):505-17.
- 175. Vekilov PG, Monaco BR, Thomas BR, Stojanoff V, Rosenberger F. Repartitioning of NaCl and protein impurities in lysozyme crystallization. Acta Crystallogr D Biol Crystallogr. 1996 Jul 1;52(Pt 4):785-98.
- 176. Vekilov PG. Protein crystal growth—microgravity aspects. *Adv Space Res.* 1999;24(10):1231-40.
- 177. Vergara A, Corvino E, Sorrentino G, Piccolo C, Tortora A, Carotenuto L, Mazzarella L, Zagari A. Crystallization of the collagen-like polypeptide (PPG)10 aboard the International Space Station. 1. Video observation. *Acta Crystallogr D Biol Crystallogr.* 2002 Oct;58(Pt 10 Pt 1):1690-4. Epub 2002 Sep 26.
- Wagner G. Bacteriorhodopsin crystal growth under microgravity—results of IML-1 and Spacehab-1 experiments. *ESA J.* 1994;18(1):25-32.
- 179. Wakayama NI, Yin DC, Harata K, Kiyoshi T, Fujiwara M, Tanimoto Y. Macromolecular crystallization in microgravity generated by a superconducting magnet. *Ann N Y Acad Sci.* 2006 Sep;1077:184-93.
- 180. Wang Y, Pan J, Niu X, Zhou Y, Hua Z, Xu Q, Wu S, Han Q, Li H, Liu Y, Teng M, He H, Lin S, Bi R. The second space experiment of protein crystallization with domestic facilities. Sci China C Life Sci. 1996 Oct;39(5):458-64.
- Wang YP, Han Y, Pan JS, Wang KY, Bi RC. Protein crystal growth in microgravity using a liquid/liquid diffusion method. *Microgravity Sci Technol.* 1996;9(4):281-3.
- Wardell MR, Skinner R, Carter DC, Twigg PD, Abrahams JP. Improved diffraction of antithrombin crystals grown in microgravity. Acta Crystallogr *D Biol Crystallogr.* 1997 Sep 1;53(Pt 5):622-5.
- 183. Wei Y, Xu G, Zhang K, Yang Z, Guo Y, Huang C, Wei B. Anomalous shear band characteristics and extra-deep shock-affected zone in Zr-based bulk metallic glass treated with nanosecond laser peening. *Sci Rep.* 2017 Mar 7;7:43948. doi: 10.1038/srep43948.
- Wilson WW, Delucas LJ. Applications of the second virial coefficient: protein crystallization and solubility. Acta Crystallogr F Struct Biol Commun. 2014 May;70(Pt 5):543-54. doi: 10.1107/S2053230X1400867X. Epub 2014 Apr 30. Erratum in: Acta Crystallogr F Struct Biol Commun. 2016 Mar 1;72(Pt 3):255-6.
- 185. Wilson, K. P., M. M. Yamashita, M. D. Sintchak, S. H. Rotstein, M. A. Murcko, J. Boger, J. A. Thomson, M. J. Fitzgibbon, J. R. Black and M. A. Navia. Comparative X-ray structures of the major binding protein for the immunosuppressant FK506 (tacrolimus) in unliganded form and in complex with FK506 and rapamycin. Acta Crystallographica Section D. 1995;51(4): 511-521.



- 186. Yau ST, Thomas BR, Galkin O, Gliko O, Vekilov PG. Molecular mechanisms of microheterogeneity-induced defect formation in ferritin crystallization. *Proteins*. 2001 Jun 1;43(4):343-52.
- Yau ST, Vekilov PG. Direct observation of nucleus structure and nucleation pathways in apoferritin crystallization. *J Am Chem Soc.* 2001 Feb 14;123(6):1080-9.
- 188. Yokoyama E, Yoshizaki I, Shimaoka T, Sone T, Kiyota T, Furukawa Y. Measurements of growth rates of an ice crystal from supercooled heavy water under microgravity conditions: basal face growth rate and tip velocity of a dendrite. *J Phys Chem B.* 2011 Jul 14;115(27):8739-45. doi: 10.1021/jp110634t. Epub 2011 Jun 20.
- 189. Yoshizaki I, Nakamura H, Fukuyama S, Komatsu H, Yoda S. Scientific approach to the optimization of protein crystallization conditions for microgravity experiments. *Ann* N Y Acad Sci. 2004 Nov;1027:28-47.
- 190. Zagalsky PF, Wright CE, Parsons M. Crystallisation of alpha-crustacyanin, the lobster carapace astaxanthinprotein: results from EURECA. *Adv Space Res.* 1995;16(8):91-4.
- Zepeda S, Nakatsubo S, Furukawa Y. Apparatus for single ice crystal growth from the melt. *Rev Sci Instrum.* 2009 Nov;80(11):115102. doi: 10.1063/1.3222739.
- 192. Zhukhovitskii DI, Fortov VE, Molotkov VI, Lipaev AM, Naumkin VN, Thomas HM, Ivlev AV, Schwabe M, Morfill GE. Nonviscous motion of a slow particle in a dust crystal under microgravity conditions. *Phys Rev E Stat Nonlin Soft Matter Phys.* 2012 Jul;86(1 Pt 2):016401. Epub 2012 Jul 9.
- 193. Zorb Ch, Weisert A, Stapelmann J, Smolik G, Carter DC, Wright BS, Brunner-Joos KD, Wagner G. Bacteriorhodopsin crystal growth in reduced gravity results under the conditions, given in CPCF on board of a Space Shuttle, versus the conditions, given in DCAM on board of the Space Station Mir. *Microgravity Sci Technol.* 2002;13(3):22-9.

